

DATABASE

Open Access



A crowdsourcing database for the copy-number variation of the Spanish population

Daniel López-López^{1,2,3}, Gema Roldán¹, Jose L. Fernández-Rueda¹, Gerrit Bostelmann¹, Rosario Carmona^{1,3}, Virginia Aquino¹, Javier Perez-Florido^{1,2}, Francisco Ortuño^{1,4}, Guillermo Pita⁵, Rocío Núñez-Torres⁵, Anna González-Neira⁵, CSVS Crowdsourcing Group, María Peña-Chilet^{1,2,3} and Joaquín Dopazo^{1,2,3,33*}

Abstract

Background Despite being a very common type of genetic variation, the distribution of copy-number variations (CNVs) in the population is still poorly understood. The knowledge of the genetic variability, especially at the level of the local population, is a critical factor for distinguishing pathogenic from non-pathogenic variation in the discovery of new disease variants.

*Correspondence:

Joaquín Dopazo

joaquin.dopazo@juntadeandalucia.es

¹ Computational Medicine Platform, Andalusian Public Foundation Progress and Health-FPS, 41013 Seville, Spain

² Institute of Biomedicine of Seville, IBI5, University Hospital Virgen del Rocío/CSIC/University of Seville, Seville, Spain

³ Centro de Investigación Biomédica en Red en Enfermedades Raras (CIBERER), ISCIII, Madrid, Spain

⁴ Department of Computer Architecture and Computer Technology, University of Granada, 18071 Granada, Spain

⁵ Human Genotyping Unit—CeGen, Spanish National Cancer Research Centre (CNIO), 28029 Madrid, Spain

⁶ Navarrabiomed-IdiSNA, Complejo Hospitalario de Navarra, IdiSNA (Navarra Institute for Health Research), Universidad Pública de Navarra (UPNA), Pamplona, Navarre, Spain

⁷ Department of Genetics, Instituto de Investigación Sanitaria-Fundación Jiménez Díaz University Hospital, Universidad Autónoma de Madrid (IIS-FJD, UAM), Madrid, Spain

⁸ Fundación Para la Investigación y Docencia Sant Joan de Deu, Barcelona, Spain

⁹ University Hospital Virgen del Rocío, Seville, Spain

¹⁰ Fundación Pública Galega de Medicina Xenómica, SERGAS, IDIS, Santiago de Compostela, Spain

¹¹ Asociación Instituto de Investigación Sanitaria de Biocruces, Vizcaya, Spain

¹² Hospital Univ. 12 de Octubre, Madrid, Spain

¹³ Centro de Investigación Príncipe Felipe, Valencia, Spain

¹⁴ Universidad de Barcelona, Barcelona, Spain

¹⁵ Hospital Virgen de la Arrixaca, Murcia, Spain

¹⁶ Servicio Madrileño de Salud, Madrid, Spain

¹⁷ Department of Genomic Medicine, Centre for Genomics and Oncological Research (GENYO), Pfizer University of Granada, Granada, Spain

¹⁸ Vall d'Hebron Institut de Recerca, Barcelona, Spain

¹⁹ Fundación para la Investigación del Hospital la Fe, Valencia, Spain

²⁰ Servicio de Genética, Ramón y Cajal Institute of Health Research (IRYCIS) and Biomedical Network Research Centre on Rare Diseases (CIBERER), Madrid, Spain

²¹ Vall d'Hebron Institut de Recerca (VHIR), Hospital Universitari Vall d'Hebron, Barcelona, Spain

²² Undiagnosed Rare Diseases Programme (ENoD), Center for Biomedical Research on Rare Diseases (CIBERER), ISCIII, Madrid, Spain

²³ Universidad de Murcia, Murcia, Spain

²⁴ Fundación IDIBELL, Barcelona, Spain

²⁵ Universidad Autónoma de Madrid, Madrid, Spain

²⁶ Universidad Pompeu Fabra, Barcelona, Spain

²⁷ Agencia Estatal Consejo Superior de Investigaciones Científicas, Madrid, Spain

²⁸ Fundación IDIBELL, Barcelona, Spain

²⁹ Universidad de Zaragoza, Saragossa, Spain

³⁰ Hospital Clínico y Provincial de Barcelona, Barcelona, Spain

³¹ Fundación Instituto de Investigación Sanitaria Illes Balears (IdISBa), Palma, Spain

³² Universidad Autónoma de Barcelona, Barcelona, Spain

³³ FPS/ELIXIR-ES, Andalusian Public Foundation Progress and Health-FPS, 41013 Seville, Spain



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Results Here, we present the SPANish Copy Number Alterations Collaborative Server (SPACNACS), which currently contains copy number variation profiles obtained from more than 400 genomes and exomes of unrelated Spanish individuals. By means of a collaborative crowdsourcing effort whole genome and whole exome sequencing data, produced by local genomic projects and for other purposes, is continuously collected. Once checked both, the Spanish ancestry and the lack of kinship with other individuals in the SPACNACS, the CNVs are inferred for these sequences and they are used to populate the database. A web interface allows querying the database with different filters that include ICD10 upper categories. This allows discarding samples from the disease under study and obtaining pseudo-control CNV profiles from the local population. We also show here additional studies on the local impact of CNVs in some phenotypes and on pharmacogenomic variants. SPACNACS can be accessed at: <http://csvs.clinbioinfospa.es/spacnacs/>.

Conclusion SPACNACS facilitates disease gene discovery by providing detailed information of the local variability of the population and exemplifies how to reuse genomic data produced for other purposes to build a local reference database.

Background

Copy-number variations (CNVs) is a frequent form of genetic variation that is increasingly being linked to genetic and phenotypic diversity as well as to disease [1–3]. The interest in the assessment of CNVs is growing among the rare diseases community in recent years, given that they can explain cases that remain unsolved after conventional single nucleotide variant (SNV) prioritization [4]. Since there is a high level of natural (and apparently non-pathogenic) CNV mutational background [5, 6], it is important to have a reference repository that provides local population context and helps to distinguish these benign CNVs from potential pathogenic CNV findings in patients. As in the case of SNV variation, the local component of CNV variation is of utmost importance [7]. However, general databases, such as DECIPHER [8] or Gnomad [9] do not contain specific data from local populations that reflect the peculiarities of their CNV variant distributions. To offer this relevant aspect to genetic studies, the SPANish Copy Number Alteration Collaborative Server (SPACNACS) stores CNV variation for more than 400 unrelated individuals of estimated Spanish ancestry. This database has been generated as a collaborative effort of the Spanish Network for Research in Rare Diseases (CIBERER) [10], the Navarra genome Project (NaGen) [11], and other research groups under a crowdsourcing cooperative model. Actually, our participation in projects like the undiagnosed patients programme (EnoD from the CIBERER) [12] guarantees a continuous submission of new patients to the database.

Since obtaining genomic data on a significant number of confirmed healthy people is often difficult, the strategy used here lies on the use of two annotations for the individuals: top levels of ICD10 and Human Phenotype Ontology (HPO) [13] annotations. This information allows building ad hoc queries in which the features of the studied individual are absent. In this way pseudo-control

cohorts can be easily constructed in which, for example, Fanconi Anemia patients can take the role of pseudo-healthy reference for patients with retinal dystrophy (and vice versa). Although pleiotropies cannot be completely ruled out, they are expected to be infrequent across ICD10 top categories or high-level HPO terms.

The availability of the local population variability at the level of CNV opens the door to additional relevant studies, such as the contribution of natural copy number variation to the pharmacogenetic profile of the Spanish population. Because of the increasing abundance of genomic data, most of the genomic variations associated with pharmacogenomics are Single Nucleotide Variants (SNVs) and small indels [14], and the pharmacogenetic profile in the Spanish population has recently been described by us [15]. However, the role of CNVs in pharmacogenomic variation remains largely unknown and cannot be ignored [16]. A clear example of the potential role of these variants is the effect of the CNVs in the *CYP2D6* gene, which encodes an enzyme which is key in the metabolism of xenobiotics, including several drugs such as opioids [17], tricyclic antidepressants [18], selective serotonin reuptake Inhibitors [19], tamoxifen [20] or ondansetron/tropisetron [21].

The SPACNACS database is an example for future federated European infrastructures [22], whose aim is to enable discovery and analysis of genomic data without having direct access to them [23]. Moreover, SPACNACS is actively participating in the CNV specifications for the new Beacon 2.0 [24] standard that will facilitate the federated analysis of genomic data at CNV level for the first time. Interestingly, SPACNACS combines the discoverability possibilities offered by a Beacon with the possibility of contacting the group that generated the sequence, a useful feature present in tools like Matchmaker Exchange [25].

Implementation

Data

A subset of high-quality genomes and exomes from the Collaborative Spanish Variant Server database (CSVS) [15] were used to populate SPACNACS, and guarantee a continuous flux of new CNV data as CSVS grows. Genomic sequences come from different projects such as the Medical Genome Project [7, 26], the EnoD, (Undiagnosed Rare Diseases programme) from the Spanish Network for Research in Rare Diseases (CIBERER), the Project Genome 1000 Navarra, The RareGenomics [27] from Madrid, and other research groups and initiatives across Spain [28–30].

Sample locality and potential kinship

The database contains only CNVs derived from unrelated individuals of Spanish ancestry. A leave-one-out cross-validation (LOOCV) strategy, previously used as a quality assessment to populate the CSVS [15], was utilized to build a distribution of percentages of variants contributed by any single sample to the pool of variants present in the rest of the database. Samples 1.5 times under the first interquartile range were considered genetically too close [15] and not included in the database. On the other hand, a Machine Learning based model, trained with different populations from the 1000 genomes project [31], was used to discriminate Spanish samples from the rest of populations (see [15] for details).

Copy number variation predictions

For each sample, FASTQC v0.11.8 [32] was used to assess quality of raw data and fastp v0.20.0 [33] was run for quality preprocessing so that clean data is provided to downstream analysis. Then, filtered sequence reads were aligned to the reference human genome build hs37d5 (hg19) by using the BWA v0.7.16a alignment tool [34]. The obtained mapped reads (BAM files) were sorted by samtools v1.11 [35] and duplicated reads were marked to mitigate biases introduced by data generation steps by means of Picard tools v2.17.3 [36]. BAM files were later analyzed in terms of QC using in-house scripts and the ngsCAT v0.1 tool [37].

Two pipelines for predicting deletions and duplications were developed. One based on Manta v1.5.0 [38] and another on Gridss v2.7.3 [39]. In both cases, the best practices recommended in the documentation were followed. Only PASS variants were kept. Both predictions are available in SPACNACS and can be selected in the Search and Selection panel (see SPACNACS functionality section below).

Pharmacogenomic analysis of the Spanish population

In order to evaluate the pharmacogenomic impact of CNVs in the Spanish population, we studied the variability of 1049 pharmacogenes involved in drug pharmacokinetics and/or drug response (Additional file 2: Table S2) described in PharmGKB database [14, 40]. We used Bedtools [41] and Pandas [42] to calculate the frequency of genes that overlap (totally or partially) with a deletion or a duplication in the Spanish population.

Gene-phenotype associations

Gene-phenotypes associations were downloaded from the Human Phenotype Ontology database [13]. Primary HPO terms for individuals were manually assigned by clinicians and experts from the corresponding genomic projects mentioned above. Statistics and plots were generated with numpy [43], Pandas [42] and matplotlib [44] libraries.

CNV annotations

SPACNACS includes an extensive annotation of CNVs with clinically relevant databases. A CNV is annotated with the features corresponding to the specific genomic region that overlaps. Such features include: (i) Clinvar database [45], a freely accessible, public archive of reports of the relationships among human variations and phenotypes, with supporting evidence. It also provides gene-disease relationships. (ii) DisGeNET [46], an exhaustive catalog of genes and variants associated with human diseases. DisGeNET integrates data from expert curated repositories, GWAS catalogs, animal models and the scientific literature. (iii) Gene Ontology Annotation (GOA) [47, 48], which contains a mixture of manual annotation and computationally assigned GO terms describing gene products. (iv) ClinGen [49], a National Institutes of Health (NIH)-funded resource that defines the clinical relevance of genes and variants for use in precision medicine and research. (v) The Human Phenotype Ontology [13], which provides a standardized vocabulary of phenotypic abnormalities encountered in human disease. In addition, HPO annotations derived from gene-phenotype links obtained from the analysis of a patient network [50]. (vi) The wgEncodeEH000322 track from UCSC [51], which provides information about the mappability of the genome. This annotation can be useful to detect false positives and biases in CNV prediction technologies/tools [52].

Statistical methods

To estimate the functional enrichment in the CNV variants, the frequency of genes that overlap (totally or partially) with a deletion or a duplication in the Gridss

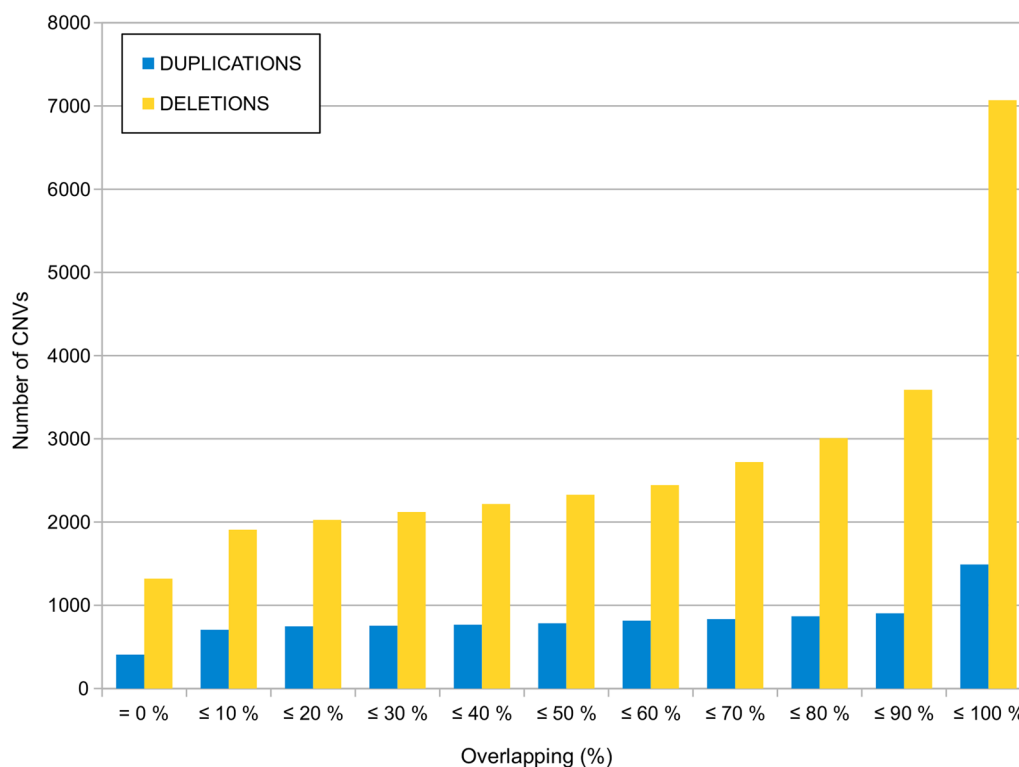


Fig. 1 CNVs overlapping between SPACNACS and other databases. Comparative between the CNVs found in SPACNACS and the ones present in the 1000 genomes and Gnomad databases. The X axis incrementally represents the level of overlap between the CNVs compared, which range from 0 (CNVs unique to SPACNACS) up to 100% (CNVs with a perfect match)

pipeline prediction dataset was first calculated. Genes affected by more than one CNV of the same individual were only counted once. Then, a z-score normalization of the frequency was performed, and those genes with a score greater or equal to four were selected. Finally, the web tool metascape [53] was used to carry out the functional enrichment.

Results

SPACNACS content description

Focusing on the Gridss pipeline, SPACNACS contains a total of 8559 unique CNVs, corresponding to 7069 deletions (83%) and 1490 duplications (17%). The reciprocal overlapping between these CNVs and those contained in two other population databases (1000 genomes project and Gnomad [9]) was evaluated. As a result, most SPACNACS CNVs overlap to a greater or lesser extent with some CNVs from both, 1000 genomes or Gnomad (Fig. 1). A total of 586 duplications (39% of the total duplications) and 3479 deletions (49% of the total deletions) of SPACNACS match almost completely with CNVs present in 1000 genomes and/or Gnomad (overlapping greater than 90%, which corresponds to the increment between the penultimate and the ultimate bars in Fig. 1).

Interestingly, a remarkable amount of SPACNACS CNVs do not overlap with any other CNV from 1000 genomes or Gnomad (see the first column of bars in Fig. 1), specifically 407 duplications (27% of the total duplications) and 1320 deletions (19% of the total deletions). These CNVs can be considered a priori either exclusive of the Spanish population or, at least, CNVs which are very abundant in the Spanish population but scarce in other populations. Therefore, these CNVs would play a crucial role in CNVs prioritization processes. The rest of CNVs display a partial overlap with CNVs present in the 1000 genomes and Gnomad databases.

From the analysis of 417 individual samples (processed with the Gridss pipeline), a total of 26,623 different sequences, corresponding to diverse biotypes affected by at least one CNV, were observed. Among them, 10,347 sequences (40%) are protein coding genes (see Additional file 1: Table S1), while the rest correspond to other biological categories (pseudogenes, RNAs, etc.). As expected, most of these genes are rarely affected by CNVs in the SPACNACS sample of the local population. Only 1064 (10.2%) coding genes are affected by at least one CNV in more than 5% of the individuals.

Since SPACNACS is composed of patients of different diseases (binned in subpopulations corresponding to the highest ICD10 levels) it is expectable that CNVs shared by several subpopulations will not be disease-specific. Given that some subpopulations can have some overlap (e.g. “5: *Mental and behavioural disorders*”, “6: *Diseases of the nervous system*”, “17: *Congenital malformations, deformations and chromosomal abnormalities*”, and the corresponding controls “31: *Mental and behavioural disorders—controls*”, “32: *Diseases of the nervous system—controls*”, “43: *Congenital malformations, deformations and chromosomal abnormalities—controls*”), a threshold of 15 different subpopulation affected have been chosen to ensure that CNVs are not disease-specific. There are 771 genes contained in CNVs that appear in individuals belonging to a total of 15 or more subpopulations. These genes constitute a conservative estimation of the genes affected by CNVs with likely no pathogenic consequences. Interestingly, 189 of them have an OMIM annotation corresponding to different inherited diseases (see Additional file 1: Table S1). This suggests that, while point mutations affecting the functionality of the gene have a pathologic effect, dose effects due to copy number alterations of the whole genes are unlikely to imply pathogenicity. Generally speaking, genes of recessive disorders would not produce any phenotype in a deletion, providing a

chromosome copy is preserved. It is also likely that a proper transcriptional control can cope with deficiencies or overabundances in the number of genes [54, 55].

Among the most frequently affected genes, there is a significant ($p\text{-value}=2.34\cdot 10^{-6}$) functional enrichment, according to the metascape tool [53] (see statistical methods section), in genes involved in *detection of chemical stimulus involved in sensory perception* (GO:0050907). This observation is in line with the fact that most olfactory receptors genes are located in segmentally duplicated regions, which are known to be frequently involved in regions affected by copy-number variation [56, 57]. Other biological functions detected in the enrichment are of more complex interpretation, given that are very general terms, such as *cell killing* (GO:0001906; $p\text{-value}=0.00065$), *single fertilization* (GO:0007338, $p\text{-value}=0.0026$), *anatomical structure maturation* (GO:0071695, $p\text{-value}=0.0068$) and *neuron projection morphogenesis* (GO:0048812, $p\text{-value}=0.0093$).

SPACNACS architecture

The SPACNACS is a web server that can be found at: <http://csvs.clinbioinfospa.es/spacnacs/>. The front-end has been developed using the JavaScript REACT library v17.0.2 [58]. The genome browser has been built using the IGV.js library [59]. The Integrative Genomics Viewer (IGV) is a popular high-performance, easy-to-use,

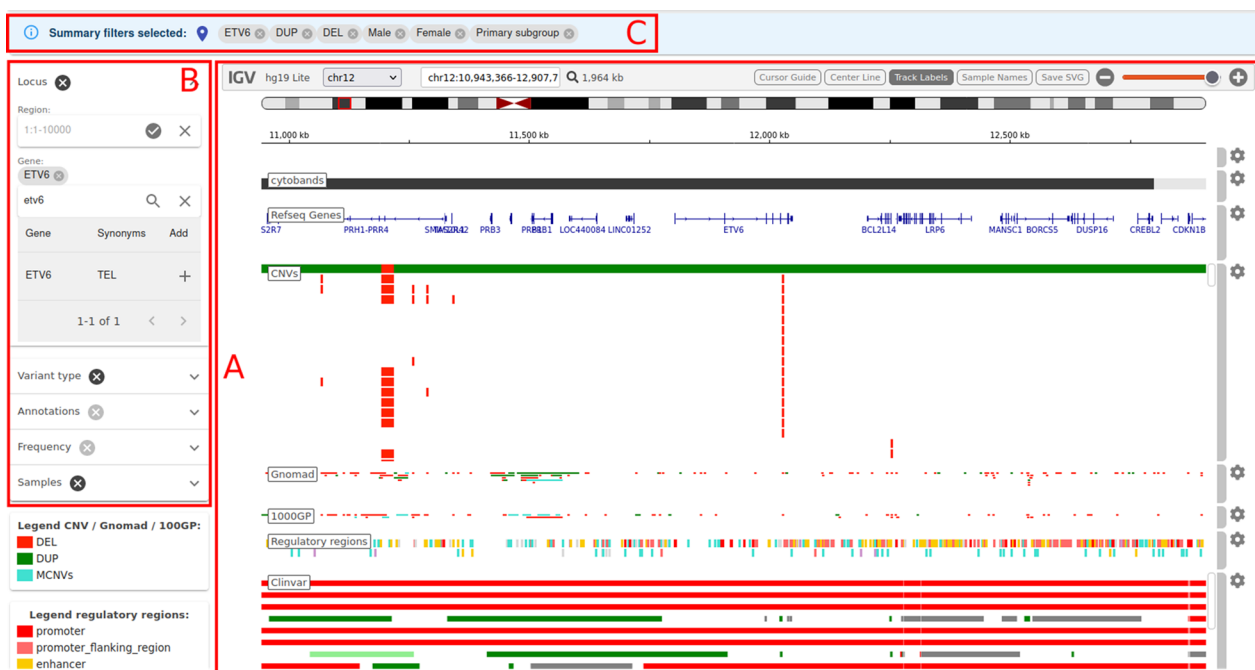


Fig. 2 The SPACNACS interface. **A** Genome browser panel consisting of an embedded Integrative Genomics Viewer preloaded with the Spanish CNV database and other useful tracks. **B** Search and selection panel, which provides several filters for specifying the genomic region and the data to be shown. **C** Filtering status panel, which shows information about the active filters. The whole dataset is shown by default

interactive tool for the visual exploration of genomic data [60]. The back-end has been written in Java programming language and the database has been built using Mongo [61], a NoSQL document database used to build highly available and scalable internet applications.

SPACNACS functionality

The web interface has 3 main sections: (i) The Browser panel (Fig. 2A), which consists of an embedded Integrative Genomics Viewer [60] preloaded with the Spanish CNV database, world population CNVs derived from Gnomad v2.1 control samples [62] and the 1000 Genomes Project structural variants phase 3 [63], regulatory regions as provided by Ensembl [64] and known clinically relevant copy number gains, copy number losses, duplications and deletions from ClinVar [45]. The browser provides several navigation controls for specifying the genomic region to view. (ii) Search and selection panel (Fig. 2B). This section controls which CNVs are displayed in the CNVs track. Several filters for specifying the genomic region and the data to be shown are provided. For example, filtering by *intellectual disability* (C3714756) term in DisGeNET will show only CNVs overlapping genes associated with this phenotype in DisGeNET. In a similar way, selecting a subset of samples according to the ICD10 category, gender or main HPO term will automatically update the CNV frequency. This allows researchers to use this repository as a pseudo-control population for ruling out non-causal CNVs and helping to find new disease-causing CNVs. In the selection panel the type of pipeline used to infer the CNVs, Manta [38] or Gridss [39] (see Methods) can be selected. (iii) Filtering status panel (Fig. 2C), which shows information about the active filters. By default, the whole dataset is selected. Additional information about the interface can be found at the server documentation main page (<https://github.com/babelomics/SPACNACS/wiki>).

The SPACNACS beacon

SPACNACS implements a Beacon (version 1.0), a standard protocol used to query the database to check whether a specific region is involved in a CNV. The Beacon is an initiative of the Global Alliance for Genomics & Health (GA4GH) that allows genomic data sharing across federated networks [65]. The Beacon is a web-accessible service that can be queried for information about one specific allele at a time. For example, in the classical Beacon a user can pose queries of the form “Have you observed this particular variation (e.g., nucleotide A) at this genomic location (e.g., position 21,926,123 on chromosome 8)?” to which the Beacon responds with either “yes” or “no.” Here, the Beacon allows queries on amplifications or deletions that involve regions. Since the

definition of the boundaries of the CNVs is often difficult, the Beacon allows querying with ranges. The generic URL to query the SPACNACS beacon is as follows: [http://csvs.clinbioinfospa.es:8080/spacnacs-ga4gh-beacon-v1/query?referenceName=\[chromosome\]&referenceBases=N&assemblyId=GRCh37&startMin=\[starMin\]&startMax=\[starMax\]&endMin=\[endMin\]&endMax=\[endMax\]&variantType=\[DUP/DEL\]](http://csvs.clinbioinfospa.es:8080/spacnacs-ga4gh-beacon-v1/query?referenceName=[chromosome]&referenceBases=N&assemblyId=GRCh37&startMin=[starMin]&startMax=[starMax]&endMin=[endMin]&endMax=[endMax]&variantType=[DUP/DEL]).

The Beacon 1.0 is used to query a region containing any base (reference Bases = *N*). For example, to search for any CNV (the & variant Type parameter is dropped) in the chromosome 8, in the locus spanning between positions 22,138,284 and 22,138,339 DEL within an interval of ± 10 nucleotides the query would be as follows: <http://csvs.clinbioinfospa.es:8080/spacnacs-ga4gh-beacon-v1/query?referenceName=8&referenceBases=N&assemblyId=GRCh37&startMin=22138273&startMax=22138293&endMin=22138328&endMax=22138348>.

SPACNACS is participating in the definition of Beacon specifications for CNV-related queries in the new Beacon 2.0 standard, currently under development [24].

Impact of CNV in human phenotype

Individuals in SPACNACS come from CSVS with detailed HPO annotations made by experts from the different genomic projects of origin. This allows carrying out an interesting correlation between phenotype and genes with phenotypic annotations in the different diseases present in the database. Table 1 lists the different HPOs found in individuals present in the database along with the HPO-related genes affected by CNVs. There are phenotypes, like *Intellectual disability*, *Global developmental delay*, *Ataxia*, *Microcephaly*, and nine more, in which all the CNVs present in affected individuals overlap genes annotated with HPOs corresponding to the phenotype. It is important to note that it does not mean that the causal genetic variation of the disease in all these individuals is a CNV. Actually, it could be the case that the diagnosis was due to a SNV for some individuals. However, it is remarkable that in some cases all the CNVs overlap, and presumably affect, genes with HPOs corresponding to the phenotype of the individual. This suggests an important role of copy number variation in some phenotypes. Actually, it is well known the role of structural variation in intellectual disabilities [66]. In other cases, like breast carcinoma, only in 10% of the patients a gene related to this HPO was affected by a deletion. In a wider spectrum of HPOs, none of the HPO-related genes was affected by a CNV. Several cancers, cardiomyopathies and retinopathies are examples of diseases typically caused by SNVs and only in an infrequent number of cases by CNVs, which agrees with the observations summarized in Table 1.

Table 1 HPOs in the individuals and HPO-related genes affected by CNVs in them

| HPO ID | HPO | Individuals | Deletions | Amplifications | Any CNV | Percentage explained |
|------------|--|-------------|-----------|----------------|---------|----------------------|
| HP:0001249 | Intellectual disability | 17 | 17 | 7 | 17 | 100.00 |
| HP:0001263 | Global developmental delay | 17 | 17 | 10 | 17 | 100.00 |
| HP:0001251 | Ataxia | 5 | 5 | 0 | 5 | 100.00 |
| HP:0000252 | Microcephaly | 4 | 4 | 3 | 4 | 100.00 |
| HP:0004322 | Short stature | 4 | 4 | 0 | 4 | 100.00 |
| HP:0001298 | Encephalopathy | 3 | 3 | 0 | 3 | 100.00 |
| HP:0002652 | Skeletal dysplasia | 2 | 2 | 0 | 2 | 100.00 |
| HP:0001256 | Intellectual disability, mild | 1 | 1 | 0 | 1 | 100.00 |
| HP:0001328 | Specific learning disability | 1 | 1 | 0 | 1 | 100.00 |
| HP:0001270 | Motor delay | 1 | 1 | 0 | 1 | 100.00 |
| HP:0001250 | Seizures | 1 | 1 | 0 | 1 | 100.00 |
| HP:0001290 | Generalized hypotonia | 1 | 1 | 1 | 1 | 100.00 |
| HP:0001067 | Neurofibromas | 1 | 1 | 0 | 1 | 100.00 |
| HP:0200134 | Epileptic encephalopathy | 4 | 3 | 0 | 3 | 75.00 |
| HP:0000729 | Autistic behavior | 2 | 1 | 0 | 1 | 50.00 |
| HP:0001332 | Dystonia | 3 | 1 | 0 | 1 | 33.33 |
| HP:0010864 | Intellectual disability, severe | 6 | 1 | 0 | 1 | 16.67 |
| HP:0003002 | Breast carcinoma | 10 | 1 | 0 | 1 | 10.00 |
| HP:0000083 | Renal insufficiency | 9 | 0 | 0 | 0 | 0.00 |
| HP:0002206 | Pulmonary fibrosis | 6 | 0 | 0 | 0 | 0.00 |
| HP:0000107 | Renal cyst | 5 | 0 | 0 | 0 | 0.00 |
| HP:0002313 | Spastic paraparesis | 5 | 0 | 0 | 0 | 0.00 |
| HP:0003003 | Colon cancer | 5 | 0 | 0 | 0 | 0.00 |
| HP:0000488 | Retinopathy | 4 | 0 | 0 | 0 | 0.00 |
| HP:0002664 | Neoplasm | 3 | 0 | 0 | 0 | 0.00 |
| HP:0002110 | Bronchiectasis | 3 | 0 | 0 | 0 | 0.00 |
| HP:0002342 | Intellectual disability, moderate | 2 | 0 | 0 | 0 | 0.00 |
| HP:0011343 | Moderate global developmental delay | 2 | 0 | 0 | 0 | 0.00 |
| HP:0009830 | Peripheral neuropathy | 2 | 0 | 0 | 0 | 0.00 |
| HP:0001300 | Parkinsonism | 2 | 0 | 0 | 0 | 0.00 |
| HP:0001258 | Spastic paraplegia | 2 | 0 | 0 | 0 | 0.00 |
| HP:0012126 | Stomach cancer | 2 | 0 | 0 | 0 | 0.00 |
| HP:0001638 | Cardiomyopathy | 2 | 0 | 0 | 0 | 0.00 |
| HP:0003107 | Abnormal circulating cholesterol concentration | 2 | 0 | 0 | 0 | 0.00 |
| HP:0004482 | Relative macrocephaly | 1 | 0 | 0 | 0 | 0.00 |
| HP:0000256 | Macrocephaly | 1 | 0 | 0 | 0 | 0.00 |
| HP:0008551 | Microtia | 1 | 0 | 0 | 0 | 0.00 |
| HP:0000525 | Abnormality iris morphology | 1 | 0 | 0 | 0 | 0.00 |
| HP:0007105 | Infantile encephalopathy | 1 | 0 | 0 | 0 | 0.00 |
| HP:0002134 | Abnormality of the basal ganglia | 1 | 0 | 0 | 0 | 0.00 |
| HP:0007002 | Motor axonal neuropathy | 1 | 0 | 0 | 0 | 0.00 |
| HP:0003477 | Peripheral axonal neuropathy | 1 | 0 | 0 | 0 | 0.00 |
| HP:0003198 | Myopathy | 1 | 0 | 0 | 0 | 0.00 |
| HP:0001324 | Muscle weakness | 1 | 0 | 0 | 0 | 0.00 |
| HP:0010978 | Abnormality of immune system physiology | 1 | 0 | 0 | 0 | 0.00 |
| HP:0100242 | Sarcoma | 1 | 0 | 0 | 0 | 0.00 |
| HP:0008527 | Congenital sensorineural hearing impairment | 1 | 0 | 0 | 0 | 0.00 |
| HP:0008504 | Moderate sensorineural hearing impairment | 1 | 0 | 0 | 0 | 0.00 |
| HP:0001639 | Hypertrophic cardiomyopathy | 1 | 0 | 0 | 0 | 0.00 |

Table 1 (continued)

| HPO ID | HPO | Individuals | Deletions | Amplifications | Any CNV | Percentage explained |
|------------|-------------------------------------|-------------|-----------|----------------|---------|----------------------|
| HP:0004356 | Abnormality of lysosomal metabolism | 1 | 0 | 0 | 0 | 0.00 |
| HP:0032245 | Abnormal metabolism | 1 | 0 | 0 | 0 | 0.00 |

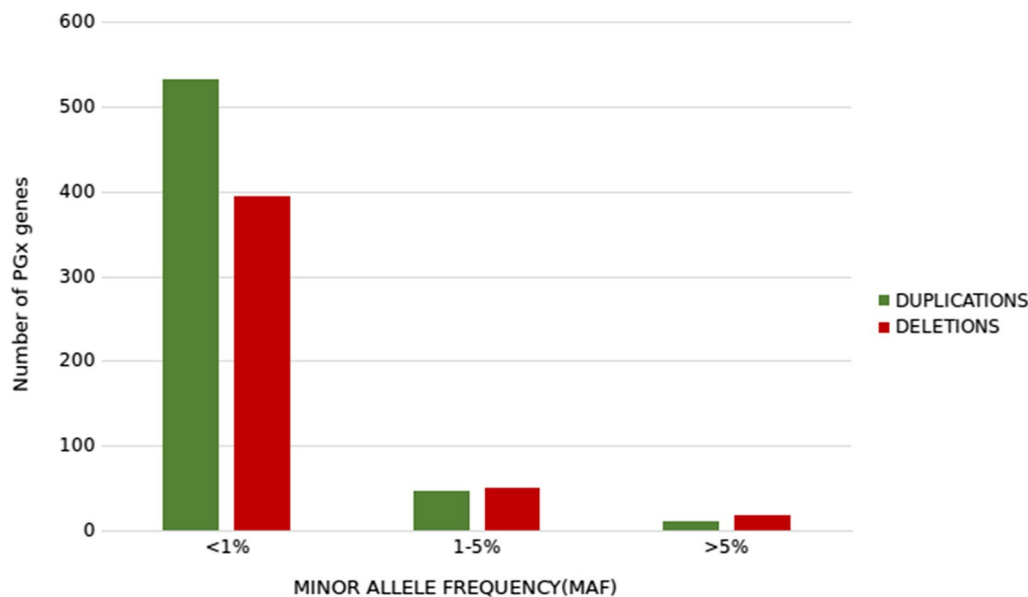


Fig. 3 Distribution of CNVs detected in pharmacogenes (PGx genes) according to the allele frequency. The number of genes harboring duplications (green) or deletions (red) of the 1045 PGx genes tested are shown according to the frequency detected in the population

Impact of CNVs on genes relevant in pharmacogenomics

In order to know the contribution of CNVs to the variability of pharmacogenomic relevant genes, the number of copies of a total of 1049 genes, reported in the clinical annotations from PharmGKB database [14], was assessed. Almost three quarters of these genes (71.7%, 749 out of 1045 pharmacogenomics genes) were involved in CNVs, with almost a half of them (49.6%, 518 out of 1045) with the full length of the gene affected by a CNV. These results document a non-negligible potential impact of CNVs on pharmacogenetic protein function. Duplications were found in a slightly more frequent proportion (56.03%) than deletions (43.97%) (Fig. 3), with 29.1% of the genes harboring both kinds of events. Of note, a 5.59% of the genes showed a minor allele frequency (MAF) higher than 5% in the Spanish population suggesting a relevant role of this kind of variation in the pharmacogenomics field. More interestingly, when the analysis was carried out only for the 21 well-described ‘actionable’ pharmacogenomics genes (PharmGKB level 1A), 80.95% of them showed CNVs with aggregated frequency of 8.29%, excluding *CYP2D6* contribution. The *CYP2D6* analysis revealed a lower CNVs frequency in

the Spanish population compared with the data reported for Caucasians (<1% versus 5.7%) [17]. These differences could be explained because the algorithm used for CNV detection is not able to solve the complexity of the structural variant rearrangements between *CYP2D6* and the pseudogene *CYP2D7* next to the gene [67–69].

Discussion

CNVs are a pervasive form of genetic variation and, as more data are collected, they are increasingly being linked to phenotypic diversity and disease [1–3, 70]. Particularly, somatic CNVs are observed in the majority of cancer types and are known to have a clear impact in cancer development and progression [71, 72]. A comprehensive representation of somatic genomic variation profiles in cancer can be found in the Progenetix database [73]. Also, numerous reports support a significant role of germinal CNVs in neurodevelopmental disorders and multiple congenital abnormalities [74, 75]. For example, more than 12% of neurodevelopmental disorder cases can be explained by a CNV [66]. It has been reported that up to 15% of the autism spectrum

could be explained by CNVs that are either de novo or rarely inherited in nature [76, 77]. Because the most characterized penetrant CNVs are inherently rare, comparative analyses against reference populations are required to assess relative disease risk and to elucidate the potential etiologic role of such genetic events, currently classified as “variants of unknown significance” (VUS) [66]. An important step in the discovery of genomic variation causal of diseases is the detailed knowledge of the local variability, as has been highlighted in the case of SNPs, where many databases with local variability have appeared in the last years [15, 78–82]. Since CNV data have traditionally been obtained at a much slower pace than other omic data, such as whole genome or whole exome sequences or transcriptomic data, finding these CNV reference population databases is often difficult. The necessity of addressing a wide range of CNV-related challenges ranging from detection and interpretation, including the lack of CNV databases with reference populations with a local component has been identified by the ELIXIR’s recently established human CNV Community [23]. Here, SPACNACS has followed the philosophy of CSVS [15], which has been described as an especially interesting example of how to collect and distribute genomic data [83], and has built a local reference of the CNV variation in the Spanish population by collecting data from different genomic projects. Another interesting feature of SPACNACS is that, instead of trying to use different tools to infer a consensus CNV profile, here, two different approaches, Manta (52) and Gridss (53) have been used and are available. Since there is not a consensus pipeline for the detection of CNVs it is interesting that people using different pipelines can find in SPACNACS different CNV estimations.

As a demonstration of the usefulness of this resource two studies using SPACNACS data are presented here. Firstly, a study of the degree of matching between the HPO annotation of individuals in SPACNACS and the corresponding HPO-related genes affected by CNVs found in them. Interestingly, the higher affectation of HPO-related genes by CNVs occurs in HPOs corresponding to diseases in which structural variation plays an important role in the etiology, such as mental disabilities or related developmental malformations [66].

Another interesting aspect is the imbalance between CNV types: deletions are significantly more frequent than duplications, a disproportion which has also been observed in other databases, such as Gnomad [9]. Actually, this imbalance is long known, as it was described that 29% of genetic diseases were caused by CNVs, being 22% of the deletions and only 7% duplications [84]. Moreover, another study in which spontaneous duplication

and deletion rates were compared to observed CNV polymorphism data from sequenced genomes, suggest that the most gene duplications are likely detrimental and are removed by natural selection [85], which can explain the observed imbalance between deletions and duplications. However, this trend is not observed in pharmacogenomic genes, which are affected almost equally by duplications (56.03%) and deletions (43.97%). Speculating the reasons for this observation is beyond the scope of this manuscript. It could be due to the fact that partial loss of function in pharmacogenomic genes is similar in deletions and duplications, contrarily to the case of essential genes mentioned above, causative of genetic diseases. Alternatively, it might be simply a matter of sampling, because the number of genes pharmacogenomic genes is not high.

Also, in spite of some limitations due to the complexity of pharmacogenomics variation, SPACNACS has demonstrated to be a valuable tool for exploring CNVs contribution in this type of genomic alteration, providing primary data about reference frequencies of pharmacogenomic genes in the Spanish population. Thus, the data presented here points to CNVs as a relevant type of variation for pharmacogenomic diagnosis and suggests their use, along with that of SVNs in the clinical implementation of pharmacogenomics.

Abbreviations

| | |
|-------|---|
| CNV | Copy number variation |
| HPO | Human phenotype ontology |
| ICD10 | International classification of diseases version 10 |
| MAF | Minor allele frequency |
| VUS | Variants of unknown significance |

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40246-023-00466-8>.

Additional file 1: Table S1. List and annotation of protein coding genes affected by at least one CNV in SPACNACS samples processed with Gridss pipeline.

Additional file 2: Table S2. List of genes involved in drug pharmacokinetics and/or drug response.

Acknowledgements

The CSVS Crowdsourcing Group: Angel Alonso⁶, Josefa Salgado-Garrido⁶, Sara Pasalodos-Sanchez⁶ (⁶Navarrabiomed-IdiSNA, Complejo Hospitalario de Navarra, Universidad Pública de Navarra (UPNA), IdiSNA (Navarra Institute for Health Research), Pamplona, Navarra, Spain). Carmen Ayuso⁷, Pablo Minguez⁷, Almudena Avila-Fernandez⁷, Marta Corton⁷ (⁷Department of Genetics, Instituto de Investigación Sanitaria-Fundación Jiménez Díaz University Hospital, Universidad Autónoma de Madrid (IIS-FJD, UAM), Madrid, Spain). Rafael Artuch⁸ (⁸Fundación para la Investigación y Docencia Sant Joan de Deu, Barcelona, Spain). Salud Borrego⁹, Guillermo Antiñolo⁹ (⁹University Hospital Virgen del Rocío, Sevilla, Spain). Angel Carracedo¹⁰, Jorge Amigo¹⁰ (¹⁰Fundación Pública Galega de Medicina Xenómica, SERGAS, IDIS, Santiago de Compostela, Spain). Luis Antonio Castaño¹¹, Isabel Tejada¹¹ (¹¹Asociación Instituto de Investigación Sanitaria de Biocruces, Vizcaya, Spain). Aitor Delmiro¹² (¹²Hospital Univ. 12 de Octubre, Madrid, Spain). Carmina Espinos¹³ (¹³Centro de

Investigación Príncipe Felipe, Valencia, Spain). Daniel Grinberg¹⁴ (¹⁴Universidad de Barcelona, Barcelona, Spain). Encarnación Guillén¹⁵ (¹⁵Hospital Virgen de la Arrixaca, Murcia, Spain). Pablo Lapunzina¹⁶ (¹⁶Servicio Madrileño de Salud, Madrid, Spain). Jose Antonio Lopez-Escámez¹⁷, Alvaro Gallego-Martinez¹⁷ (¹⁷Department of Genomic Medicine, Centre for Genomics and Oncological Research (GENYO), Pfizer University of Granada, Granada, Spain). Ramón Martí¹⁸, Eulalia Rovira¹⁸ (¹⁸Vall d'Hebron Institut de Recerca, Barcelona, Spain). José M^a Millán¹⁹ (¹⁹Fundación para la Investigación del Hospital la Fe, Valencia, Spain). Miguel Angel Moreno²⁰, Matías Morin²⁰ (²⁰Servicio de Genética, Ramón y Cajal Institute of Health Research (IRYCIS) and Biomedical Network Research Centre on Rare Diseases (CIBERER), Madrid, Spain). Antonio Moreno-Galdó²¹, Mónica Fernández-Cancio²¹ (²¹Vall d'Hebron Institut de Recerca (VHIR), Hospital Universitari Vall d'Hebron, Barcelona, Spain). Beatriz Morte²² (²²Undiagnosed Rare Diseases Programme (ENoD), Center for Biomedical Research on Rare Diseases (CIBERER), ISCIII, Madrid, Spain). Victoriano Mulero²³, Diana García²³ (²³Universidad de Murcia, Murcia, Spain). Virginia Nunes²⁴ (²⁴Fundación IDIBELL, Barcelona Spain). Francesc Palau²⁵ (²⁵Fundación para la Investigación y Docencia Sant Joan de Deu, Barcelona, Spain). Belén Perez²⁶ (²⁶Universidad Autónoma de Madrid, Madrid, Spain). Luis Pérez Jurado²⁷ (²⁷Universidad Pompeu Fabra, Barcelona, Spain). Rosario Perona²⁸ (²⁸Agencia Estatal Consejo Superior de Investigaciones Científicas, Madrid, Spain). Aurora Pujol²⁹ (²⁹Fundación IDIBELL, Barcelona, Spain). Feliciano Ramos³⁰, Esther Lopez³⁰ (³⁰Universidad de Zaragoza, Zaragoza, Spain). Antonia Ribes³¹ (³¹Hospital Clínico y Provincial de Barcelona, Barcelona, Spain). Jordi Rosell³² (³²Fundación Instituto de Investigación Sanitaria Illes Balears (IdISBa), Palma de Mallorca, Spain). Jordi Surrallés³³ (³³Universidad Autónoma de Barcelona, Barcelona, Spain).

Author contributions

DLL generate the CNV data and participated in the design of the software. GR and JLFR developed the software. GB processed the data. RC, VA and MPC analyzed and interpreted the data. JPF and FO developed the tests applied for data quality control. GP, RN and AGN carried out the pharmacogenomic analysis. The CSVS crowdsourcing group provided the original data. JD conceived the work and wrote the paper. All authors read and approved the final manuscript.

Funding

This work is supported by Grants PID2020-117979RB-I00 from the Spanish Ministry of Science and Innovation; by the Institute of Health Carlos III (project IMPaCT-Data, exp. IMP/00019, IMP/00009 and PI20/01305), co-funded by the European Union, European Regional Development Fund (ERDF, "A way to make Europe").

Availability of data and materials

Project name: SPACNACS. Project home page: <http://csvs.clinbioinfospa.es/spacnacs>. Operating system: Platform independent. Programming language: Javascript, Java and MONGO database. License: freely accessible web server.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Received: 12 September 2022 Accepted: 25 February 2023

Published online: 09 March 2023

References

- Beckmann JS, Estivill X, Antonarakis SE. Copy number variants and genetic traits: closer to the resolution of phenotypic to genotypic variability. *Nat Rev Genet.* 2007;8(8):639–46.

- Buchanan JA, Scherer SW. Contemplating effects of genomic structural variation. *Genet Med.* 2008;10(9):639–47.
- Lee C, Scherer SW. The clinical context of copy number variation in the human genome. *Exp Rev Mol Med.* 2010;12:e08.
- Royer-Bertrand B, Cisarova K, Niel-Butschi F, Mittaz-Crettol L, Fodstad H, Superti-Furga A. CNV detection from exome sequencing data in routine diagnostics of rare genetic disorders: opportunities and limitations. *Genes.* 2021;12(9):1427.
- Sudmant PH, Rausch T, Gardner EJ, Handsaker RE, Abyzov A, Huddleston J, Zhang Y, Ye K, Jun G, Hsi-Yang Fritz M. An integrated map of structural variation in 2504 human genomes. *Nature.* 2015;526(7571):75–81.
- Conrad DF, Pinto D, Redon R, Feuk L, Gokcumen O, Zhang Y, Aerts J, Andrews TD, Barnes C, Campbell P. Origins and functional impact of copy number variation in the human genome. *Nature.* 2010;464(7289):704–12.
- Dopazo J, Amadoz A, Bleda M, Garcia-Alonso L, Alemán A, García-García F, Rodriguez JA, Daub JT, Muntané G, Rueda A, et al. 267 Spanish exomes reveal population-specific differences in disease-related genetic variation. *Mol Biol Evol.* 2016;33(5):1205–18.
- Bragin E, Chatzimichali EA, Wright CF, Hurler ME, Firth HV, Bevan AP, Swaminathan GJ. DECIPHER: database for the interpretation of phenotype-linked plausibly pathogenic sequence and copy-number variation. *Nucleic Acids Res.* 2014;42(D1):D993–1000.
- Collins RL, Brand H, Karczewski KJ, Zhao X, Alföldi J, Francioli LC, Khera AV, Lowther C, Gauthier LD, Wang H, et al. A structural variation reference for medical and population genetics. *Nature.* 2020;581(7809):444–51.
- CIBERER: The Spanish Network for Research in Rare diseases. <https://www.ciberer.es/en>.
- NAGEN. Proyecto 1000 genomas de Navarra. <https://www.nagen1000navarra.es/>.
- EnoD: Undiagnosed Rare Diseases Programme. <https://www.ciberer.es/en/transversal-programmes/scientific-projects/undiagnosed-rare-diseases-programme-enod>.
- Köhler S, Gargano M, Matentzoglou N, Carmody LC, Lewis-Smith D, Vasilevsky NA, Danis D, Balagura G, Baynam G, Brower AM. The human phenotype ontology in 2021. *Nucleic Acids Res.* 2021;49(D1):D1207–17.
- Barbarino JM, Whirl-Carrillo M, Altman RB, Klein TE. PharmGKB: A world-wide resource for pharmacogenomic information. *Wiley Interdiscip Rev Syst Biol Med.* 2018;10(4): e1417.
- Peña-Chilet M, Roldán G, Perez-Florido J, Ortuño FM, Carmona R, Aquino V, Lopez-Lopez D, Loucera C, Fernandez-Rueda JL, Gallego A, et al. CSVS, a crowdsourcing database of the Spanish population genetic variability. *Nucleic Acids Res.* 2020;49(D1):D1130–7.
- He Y, Hoskins JM, McLeod HL. Copy number variants in pharmacogenetic genes. *Trends Mol Med.* 2011;17(5):244–51.
- Crews KR, Monte AA, Huddart R, Caudle KE, Kharasch ED, Gaedigk A, Dunnenberger HM, Leeder JS, Callaghan JT, Samer CF. Clinical Pharmacogenetics Implementation Consortium guideline for CYP2D6, OPRM1, and COMT genotypes and select opioid therapy. *Clin Pharmacol Ther.* 2021;110(4):888–96.
- Hicks JK, Sangkuhl K, Swen JJ, Ellingrod VL, Müller DJ, Shimoda K, Bishop JR, Kharasch ED, Skaar TC, Gaedigk A. Clinical Pharmacogenetics Implementation Consortium guideline (CPIC[®]) for CYP2D6 and CYP2C19 genotypes and dosing of tricyclic antidepressants: 2016 update. *Clin Pharmacol Ther.* 2017;102(1):37.
- Hicks JK, Bishop JR, Sangkuhl K, Müller DJ, Ji Y, Leckband SG, Leeder JS, Graham RL, Chiulli DL, Llerena A. Clinical Pharmacogenetics Implementation Consortium (CPIC) guideline for CYP2D6 and CYP2C19 genotypes and dosing of selective serotonin reuptake inhibitors. *Clin Pharmacol Ther.* 2015;98(2):127–34.
- Goetz MP, Sangkuhl K, Guchelaar HJ, Schwab M, Province M, Whirl-Carrillo M, Symmans WF, McLeod HL, Ratain MJ, Zembutsu H. Clinical Pharmacogenetics Implementation Consortium (CPIC) guideline for CYP2D6 and tamoxifen therapy. *Clin Pharmacol Ther.* 2018;103(5):770–7.
- Bell GC, Caudle KE, Whirl-Carrillo M, Gordon RJ, Hikino K, Prows CA, Gaedigk A, Agundez JA, Sadhasivam S, Klein TE. Clinical Pharmacogenetics Implementation Consortium (CPIC) guideline for CYP2D6 genotype and use of ondansetron and tropisetron. *Clin Pharmacol Ther.* 2017;102(2):213.

22. Saunders G, Baudis M, Becker R, Beltran S, Bérout C, Birney E, Brooks-bank C, Brunak S, Van den Bulcke M, Drysdale R. Leveraging European infrastructures to access 1 million human genomes by 2022. *Nat Rev Genet.* 2019;20(11):693–701.
23. Salgado D, Armean IM, Baudis M, Beltran S, Capella-Gutierrez S, Carvalho-Silva D, Del Angel VD, Dopazo J, Furlong LI, Gao B. The ELIXIR human copy number variations community: building bioinformatics infrastructure for research. *FRsearch.* 2020;9:ELIXIR-1229.
24. Rambla J, Baudis M, Ariosa R, Beck T, Fromont LA, Navarro A, Paloots R, Rueda M, Saunders G, Singh B, et al. Beacon v2 and Beacon networks: a “lingua franca” for federated data discovery in biomedical genomics, and beyond. *Hum Mutat.* 2022;43(6):791–9.
25. Philippakis AA, Azzariti DR, Beltran S, Brookes AJ, Brownstein CA, Brudno M, Brunner HG, Buske OJ, Carey K, Doll C. The matchmaker exchange: a platform for rare disease gene discovery. *Hum Mutat.* 2015;36(10):915–21.
26. The Medical Genome Project. <http://www.medicalgenomeproject.com/>.
27. References: Kindly. <https://www.rare-genomics.com>.
28. Gui H, Schriemer D, Cheng WW, Chauhan RK, Antiñolo G, Berríos C, Bleda M, Brooks AS, Brouwer RW, Burns AJ. Whole exome sequencing coupled with unbiased functional analysis reveals new Hirschsprung disease genes. *Genome Biol.* 2017;18(1):1–13.
29. Gallego-Martínez A, Lopez-Escamez JA. Genetic architecture of Meniere’s disease. *Hear Res.* 2020;397: 107872.
30. Torrent-Vernetta A, Gaboli M, Castillo-Corullón S, Mondéjar-López P, Sanz Santiago V, Costa-Colomer J, Osona B, Torres-Borrego J, de la Serna-Blázquez O, Bellón Alonso S, et al. Incidence and prevalence of children’s diffuse lung disease in Spain. *Arch Bronconeumol.* 2022;58(1):22–9.
31. Abecasis GR, Auton A, Brooks LD, DePristo MA, Durbin RM, Handsaker RE, Kang HM, Marth GT, McVean GA. An integrated map of genetic variation from 1092 human genomes. *Nature.* 2012;491(7422):56–65.
32. FastQC. A quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>.
33. Chen S, Zhou Y, Chen Y, Gu J. fastp: an ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics.* 2018;34(17):i884–90.
34. Li H, Durbin R. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics.* 2009;25(14):1754–60.
35. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R. The sequence alignment/map format and SAMtools. *Bioinformatics.* 2009;25(16):2078–9.
36. Picard. A set of command line tools (in Java) for manipulating high-throughput sequencing (HTS) data and formats such as SAM/BAM/CRAM and VCF. <http://broadinstitute.github.io/picard/>.
37. Lopez-Domingo FJ, Florido JP, Rueda A, Dopazo J, Santoyo-Lopez J. ngsCAT: a tool to assess the efficiency of targeted enrichment sequencing. *Bioinformatics.* 2014;30(12):1767–8.
38. Chen X, Schulz-Trieglaff O, Shaw R, Barnes B, Schlesinger F, Källberg M, Cox AJ, Kruglyak S, Saunders CT. Manta: rapid detection of structural variants and indels for germline and cancer sequencing applications. *Bioinformatics.* 2016;32(8):1220–2.
39. Cameron DL, Schröder J, Penington JS, Do H, Molania R, Dobrovic A, Speed TP, Papenfuss AT. GRIDSS: sensitive and specific genomic rearrangement detection using positional de Bruijn graph assembly. *Genome Res.* 2017;27(12):2050–60.
40. Whirl-Carrillo M, Huddart R, Gong L, Sangkuhl K, Thorn CF, Whaley R, Klein TE. An evidence-based framework for evaluating pharmacogenomics knowledge for personalized medicine. *Clin Pharmacol Ther.* 2021;110(3):563–72.
41. Quinlan AR. BEDTools: the swiss-army tool for genome feature analysis. *Curr Protoc Bioinform.* 2014;47(1):11.12.11–11.12.34.
42. McKinney W: Data structures for statistical computing in python. In: *Proceedings of the 9th Python in Science Conference*, pp. 51–56. Austin, TX (2010).
43. Harris CR, Millman KJ, van der Walt SJ, Gommers R, Virtanen P, Cournapeau D, Wieser E, Taylor J, Berg S, Smith NJ. Array programming with NumPy. *Nature.* 2020;585(7825):357–62.
44. Hunter JD. Matplotlib: a 2D graphics environment. *Comput Sci Eng.* 2007;9(03):90–5.
45. Landrum MJ, Lee JM, Benson M, Brown GR, Chao C, Chitipiralla S, Gu B, Hart J, Hoffman D, Jang W, et al. ClinVar: improving access to variant interpretations and supporting evidence. *Nucleic Acids Res.* 2017;46(D1):D1062–7.
46. Piñero J, Queralt-Rosinach N, Bravo À, Deu-Pons J, Bauer-Mehren A, Baron M, Sanz F, Furlong LI. DisGeNET: a discovery platform for the dynamical exploration of human diseases and their genes. *Database.* 2015;2015:bav028.
47. Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, et al. Gene ontology: tool for the unification of biology. *Gene Ontol Consort Nat Genet.* 2000;25(1):25–9.
48. Consortium GO. The gene ontology resource: enriching a gold mine. *Nucleic Acids Res.* 2021;49(D1):D325–34.
49. Rehm HL, Berg JS, Brooks LD, Bustamante CD, Evans JP, Landrum MJ, Ledbetter DH, Maglott DR, Martin CL, Nussbaum RL. ClinGen—the clinical genome resource. *N Engl J Med.* 2015;372(23):2235–42.
50. Rojano E, Seoane P, Bueno-Amoros A, Perkins JR, Garcia-Ranea JA: Revealing the relationship between human genome regions and pathological phenotypes through network analysis. In: *International conference on bioinformatics and biomedical engineering*, pp. 197–207. Springer (2017).
51. Rosenbloom KR, Sloan CA, Malladi VS, Dreszer TR, Learned K, Kirkup VM, Wong MC, Maddren M, Fang R, Heitner SG. ENCODE data in the UCSC genome browser: year 5 update. *Nucleic Acids Res.* 2012;41(D1):D56–63.
52. Derrien T, Estellé J, Marco Sola S, Knowles DG, Raineri E, Guigó R, Ribeca P. Fast computation and applications of genome mappability. *PLoS ONE.* 2012;7(1): e30377.
53. Zhou Y, Zhou B, Pache L, Chang M, Khodabakhshi AH, Tanaseichuk O, Benner C, Chanda SK. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nat Commun.* 2019;10(1):1–10.
54. The Wellcome Trust Case Control Consortium. Genome-wide association study of CNVs in 16,000 cases of eight common diseases and 3,000 shared controls. *Nature.* 2010;464(7289):713–20.
55. Gamazon ER, Stranger BE. The impact of human copy number variation on gene expression. *Brief Funct Genomics.* 2015;14(5):352–7.
56. Young JM, Endicott RM, Parghi SS, Walker M, Kidd JM, Trask BJ. Extensive copy-number variation of the human olfactory receptor gene family. *Am J Human Genetics.* 2008;83(2):228–42.
57. Cooper GM, Nickerson DA, Eichler EE. Mutational and selective effects on copy-number variants in the human genome. *Nat Genet.* 2007;39(7):S22–9.
58. REACT library. <https://reactjs.org/>.
59. IGV. <https://software.broadinstitute.org/software/igv>.
60. Robinson JT, Thorvaldsdóttir H, Winckler W, Guttman M, Lander ES, Getz G, Mesirov JP. Integrative genomics viewer. *Nat Biotechnol.* 2011;29(1):24–6.
61. MONGODB database. <https://www.mongodb.com/atlas/database>.
62. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, Collins RL, Laricchia KM, Ganna A, Birnbaum DP. The mutational constraint spectrum quantified from variation in 141,456 humans. *Nature.* 2020;581(7809):434–43.
63. Genomes Project Consortium. A global reference for human genetic variation. *Nature.* 2015;526(7571):68.
64. Cunningham F, Allen JE, Allen J, Alvarez-Jarreta J, Amode MR, Armean Irina M, Austine-Orimoloye O, Azov Andrew G, Barnes I, Bennett R, et al. Ensembl 2022. *Nucleic Acids Res.* 2021;50(D1):D988–95.
65. Fiume M, Cupak M, Keenan S, Rambla J, de la Torre S, Dyke SOM, Brookes AJ, Carey K, Lloyd D, Goodhand P, et al. Federated discovery and sharing of genomic data using Beacons. *Nat Biotechnol.* 2019;37(3):220–4.
66. Miller DT, Adam MP, Aradhya S, Biasecker LG, Brothman AR, Carter NP, Church DM, Crolla JA, Eichler EE, Epstein CJ. Consensus statement: chromosomal microarray is a first-tier clinical diagnostic test for individuals with developmental disabilities or congenital anomalies. *Am J Human Genet.* 2010;86(5):749–64.
67. Gaedigk A, Ingelman-Sundberg M, Miller NA, Leeder JS, Whirl-Carrillo M, Klein TE, Committee PS. The Pharmacogene Variation (PharmVar) Consortium: incorporation of the human cytochrome P450 (CYP) allele nomenclature database. *Clin Pharmacol Ther.* 2018;103(3):399–401.
68. Chen X, Shen F, Gonzaludo N, Malhotra A, Rogert C, Taft RJ, Bentley DR, Eberle MA. Cyrius: accurate CYP2D6 genotyping using whole-genome sequencing data. *Pharmacogenomics J.* 2021;21(2):251–61.
69. Twesigomwe D, Drögemöller BI, Wright GE, Siddiqui A, da Rocha J, Lombard Z, Hazelhurst S. StellarPGx: a nextflow pipeline for calling star alleles in cytochrome P450 genes. *Clin Pharmacol Ther.* 2021;110(3):741–9.

70. Uddin M, Thiruvahindrapuram B, Walker S, Wang Z, Hu P, Lamoureux S, Wei J, MacDonald JR, Pellicchia G, Lu C. A high-resolution copy-number variation resource for clinical and population genetics. *Genet Med*. 2015;17(9):747–52.
71. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646–74.
72. Albertson DG, Collins C, McCormick F, Gray JW. Chromosome aberrations in solid tumors. *Nat Genet*. 2003;34(4):369–76.
73. Huang Q, Carrio-Cordo P, Gao B, Paloots R, Baudis M. The Progenetix oncogenomic resource in 2021. *Database*. 2021;2021:baab043.
74. Cook EH Jr, Scherer SW. Copy-number variations associated with neuropsychiatric conditions. *Nature*. 2008;455(7215):919–23.
75. Pinto D, Pagnamenta AT, Klei L, Anney R, Merico D, Regan R, Conroy J, Magalhaes TR, Correia C, Abrahams BS. Functional impact of global rare copy number variation in autism spectrum disorders. *Nature*. 2010;466(7304):368–72.
76. Devlin B, Scherer SW. Genetic architecture in autism spectrum disorder. *Curr Opin Genet Dev*. 2012;22(3):229–37.
77. Stobbe G, Liu Y, Wu R, Hudgings LH, Thompson O, Hisama FM. Diagnostic yield of array comparative genomic hybridization in adults with autism spectrum disorders. *Genet Med*. 2014;16(1):70–7.
78. Fattahi Z, Beheshtian M, Mohseni M, Poustchi H, Sellars E, Nezhadi SH, Amini A, Arzhanghi S, Jalalvand K, Jamali P. Iranome: a catalog of genomic variations in the Iranian population. *Hum Mutat*. 2019;40(11):1968–84.
79. Nagasaki M, Yasuda J, Katsuoka F, Nariai N, Kojima K, Kawai Y, Yamaguchi-Kabata Y, Yokozawa J, Danjoh I, Saito S, et al. Rare variant discovery by deep whole-genome sequencing of 1070 Japanese individuals. *Nat Commun*. 2015;6:8018.
80. Gudbjartsson DF, Helgason H, Gudjonsson SA, Zink F, Oddson A, Gylfason A, Besenbacher S, Magnusson G, Halldorrsson BV, Hjartarson E, et al. Large-scale whole-genome sequencing of the Icelandic population. *Nat Genet*. 2015;47(5):435–44.
81. The_Genome_of_the_Netherlands_Consortium: Whole-genome sequence variation, population structure and demographic history of the Dutch population. *Nat Genet*. 2014;46(8):818–825.
82. Lim ET, Wurtz P, Havulinna AS, Palta P, Tukiainen T, Rehnstrom K, Esko T, Magi R, Inouye M, Lappalainen T, et al. Distribution and medical impact of loss-of-function variants in the Finnish founder population. *PLoS Genet*. 2014;10(7): e1004494.
83. Wojcik GL, Murphy J, Edelson JL, Gignoux CR, Ioannidis AG, Manning A, Rivas MA, Buyske S, Hendricks AE. Opportunities and challenges for the use of common controls in sequencing studies. *Nat Rev Genet*. 2022;23(11):665–79.
84. Botstein D, Risch N. Discovering genotypes underlying human phenotypes: past successes for Mendelian disease, future approaches for complex disease. *Nat Genet*. 2003;33(3):228–37.
85. Katju V, Bergthorsson U. Copy-number changes in evolution: rates, fitness effects and adaptive significance. *Front Genet*. 2013;4:273.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

