

The impact of sample size and marker selection on the study of haplotype structures

Xiao Sun,^{1,2} J. Claiborne Stephens¹ and Hongyu Zhao^{2*}

¹Genaissance Pharmaceuticals, 5 Science Park, New Haven, CT 06511, USA

²Yale University School of Medicine, 60 College Street, New Haven, CT 06520, USA

*Correspondence to: Tel: +1 203 785 6271; Fax: +1 203 785 6912, E-mail: hongyu.zhao@yale.edu

Date received (in revised form): 22nd December 2003

Abstract

Several studies of haplotype structures in the human genome in various populations have found that the human chromosomes are structured such that each chromosome can be divided into many blocks, within which there is limited haplotype diversity. In addition, only a few genetic markers in a putative block are needed to capture most of the diversity within a block. There has been no systematic empirical study of the effects of sample size and marker set on the identified block structures and representative marker sets, however. The purpose of this study was to conduct a detailed empirical study to examine such impacts. Towards this goal, we have analysed three representative autosomal regions from a large genome-wide study of haplotypes with samples consisting of African-Americans and samples consisting of Japanese and Chinese individuals. For both populations, we have found that the sample size and marker set have significant impact on the number of blocks and the total number of representative markers identified. The marker set in particular has very strong impacts, and our results indicate that the marker density in the original datasets may not be adequate to allow a meaningful characterisation of haplotype structures. In general, we conclude that we need a relatively large sample size and a very dense marker panel in the study of haplotype structures in human populations.

Keywords: single nucleotide polymorphism (SNP), haplotype, sample size, marker selection, haplotype block, tag SNPs

Introduction

Human DNA sequence variation accounts for a large fraction of the observed phenotypic differences between individuals, including susceptibility to disease. Sites in the DNA sequence where individuals differ at a single DNA base are called single nucleotide polymorphisms (SNPs). SNPs represent by far the most common source of genetic variation, and it is estimated that the human genome may contain over 10 million SNPs, about one in every 300 bases.^{1–3}

A haplotype is the specific combination of marker alleles within a region of a chromosome. Tightly linked SNPs are not independent on a given chromosome, but tend to be associated with each other across small regions. This tendency is called linkage disequilibrium. Empirical data suggest that relatively few of the theoretically possible haplotypes are observed at significant frequencies for a set of SNPs within a very short physical distance.⁴

Genome-wide disease association studies using SNPs and haplotypes may be the most promising approach to identifying genetic variants underlying complex diseases, and recent

technological advances have made high-throughput sequencing and genotyping possible. With the aim of speeding the discovery of genes related to common illnesses, as well as preventing adverse drug reactions, the National Institutes of Health launched the international HapMap Project to organise what is known about genetic variation within the human genome. One objective of this project was to understand haplotype structures throughout the human genome.

Recent studies^{5–8} have shown that haplotypes may be divided into discrete blocks, within which there is limited haplotype diversity. For example, Gabriel and colleagues systematically examined 51 autosomal regions in four populations and found that the minimal span of the blocks averaged 9 kb in Yoruban and African-American samples, with a range of <1 kb up to 94 kb, whereas the average in European and Asian samples was 18 kb, with a range of <1 kb to 73 kb.⁸ Furthermore, in a study of the class II region of the major histocompatibility complex, researchers found that the haplotype blocks were flanked by precisely localised recombination hotspots, leading to the hypothesis that ‘punctuate recombination’ could be the molecular mechanism underlying block structure.⁹

One attractive feature of statistical association methods based on haplotype blocks is the idea that, although blocks may contain a large number of SNPs, only a few SNPs are needed to uniquely identify the haplotypes in a block. This much smaller subset of SNPs, which are termed 'haplotype tagging SNPs' (htSNPs), can be used to explain a large proportion of diversity. Tag SNPs make it unnecessary to genotype all the SNPs in a given region and therefore represent an economic approach to genome-wide association studies. Zhang *et al.*¹⁰ studied the power of different association tests in a variety of disease models by using Tag SNPs and concluded that the genotyping efforts can be significantly reduced without much loss of power.

Despite these findings of block-like structures in the human genome, there is no universally accepted definition of haplotype blocks. In fact, each study has its own definition. Different definitions of haplotype blocks include: (1) a continuous set of markers in which the average pairwise D' is greater than some predetermined threshold;¹¹ (2) a region where a small number of common haplotypes account for the majority of the chromosomes;^{6,12} (3) regions with both limited haplotype diversity and strong linkage disequilibrium but allowing several markers to be skipped;⁷ and (4) regions with absolutely no evidence for historical recombination between any pair of SNPs.¹³ Therefore, block definition remains subjective and arbitrary, and it is not yet clear how to compare haplotype blocks between studies. Furthermore, each method varies in terms of the SNP minor allele frequency threshold used. The most appropriate definition may depend on how the inferred blocks are used, such as whether the identified blocks will be used to infer recombination hot spots or to identify regions that are associated with disease. Moreover, recent studies suggest that there may be non-trivial departures from block structures.¹⁴

Despite extensive empirical studies on haplotype blocks, one issue that has not been well addressed is the impact of sample size on the assessment of haplotype block structure. In some previous studies, blocks were identified based on a small set of chromosomes and may not provide a comprehensive representation of the whole population. For example, the chromosome 21 study only examined 20 independent chromosomes from diverse populations.⁶ The largest dataset reported to date contains samples from 275 individuals, leading to 400 independent chromosomes.⁸ It is not known, however, how many individuals are sufficient to get reliable characterisation of haplotype block structures.

In addition, the effect of SNP marker selection on the inferred haplotype block structures has not been well studied either. To date, the density of SNPs analysed has ranged from approximately one marker per kilobase^{6,9} to one marker per 15 kb.⁷ Published results suggest that a denser marker panel tends to give rise to a larger number of shorter blocks,⁶ whereas a sparser marker panel generates fewer longer blocks.^{7,8} Furthermore, the block boundaries and Tag SNPs

may substantially change, even if we keep the SNP density constant but select a different set of SNPs. In a recent study by Wall and Pritchard,¹⁵ they found using simulations that marker density is more important than sample size for inferring haplotype structures.

One of the objectives of the HapMap project is to understand population differences in their haplotype structures. It is important to compare haplotype blocks in different populations and to examine whether the same set of Tag SNPs can be used in different populations to capture haplotype diversities. Existing data have shown that the blocks in a Yoruban population from Nigeria are generally the same as, but shorter than, those in European and Asian populations.⁸ If different populations indeed share similar haplotype block structures, one broad map would be sufficient. If the populations are different enough, however, it might be necessary to construct population-specific haplotype maps.

These are very important questions requiring answers, and the data collected from the HapMap project may help us to gain a better understanding of these issues. In the current study, we focused on the impact of sample size and SNP marker selection on the haplotype block partitioning and Tag SNP selections in a sample consisting of African-Americans and a sample consisting of Japanese and Chinese people.

Materials and methods

Datasets

SNP genotype data of 51 autosomal regions that collectively span ~0.4 per cent of the human genome from African-American samples (called population B in the original study) and from Japanese and Chinese samples (called population C in the original study) were downloaded from the following website: <http://www.genome.wi.mit.edu/mpg/hapmap/hapstruc.html>. A detailed description of the data can be found in the paper by Gabriel *et al.*⁸ Population B contains 50 samples from unrelated African-Americans and population C includes 42 samples from unrelated individuals of Japanese and Chinese origin. This is the largest public dataset available to date.

In order to examine the impact of sample size and marker selection on haplotype block boundaries and Tag SNPs, three regions from the above database were chosen in our study. Region 52a spans 237.22 kb on chromosome 22 and contains 46 SNPs for population B and 45 SNPs for population C. Region 42a is 409.92 kb long and is located on chromosome 15, it includes 100 SNPs for population B and 99 SNPs for population C. Region 31a is the shortest of the three. It is on chromosome 9, is 181.98 kb long and has 23 SNPs for population B and 25 SNPs for population C. The density of the markers in these three regions is one SNP per 4 to 8 kb. We chose these three regions because they represent small, medium and large numbers of SNPs within a given region in this dataset.

Haplotype block partitioning and Tag SNP selections

To obtain haplotype boundaries and Tag SNPs, we used 'HapBlock', a dynamic programming algorithm for haplotype block partitioning with minimum number of Tag SNPs developed by Zhang *et al.*¹² The following parameters were used in our analysis: the input data type was genotype data; the method for block definition was the one used in Patil *et al.*,⁶ the threshold to define the block was set at 0.8; the threshold to define the common haplotype was set at 0.099; the method to find the Tag SNPs was the haplotype block diversity introduced by Johnson *et al.*;¹⁶ and the threshold to find the Tag SNPs was set at 0.9.

Impact of sample size

To examine the impact of sample size on the identified haplotype structures, we randomly selected 10, 20, 30 and 40 individuals out of 50 African-Americans in population B and repeated the random selection 100 times. For each randomly selected sample, we took their SNP genotype data in regions 52a, 42a and 31a and ran the HapBlock program to identify the number of blocks, the block boundaries and the Tag SNPs for each block. The same procedures were applied to population C, which included 42 unrelated Japanese and Chinese people. These results were used to assess the effect of sample size on haplotype block structures.

Impact of marker selection

Random marker selection. To study the impact of marker selection on the assessment of haplotype block structures, we carried out random selection on SNP markers for the three regions. Because region 52a contains 46 SNPs for population B (African-American) and 45 SNPs for population C (Japanese and Chinese), we randomly selected 10, 20, 30 and 40 SNPs for each population and repeated random selection 100 times. For region 42a, which includes 100 SNPs for population B and 99 SNPs for population C, we randomly selected 20, 40, 60 and 80 SNPs for each population and repeated this 100 times. Similarly for region 31a, where there are 23 SNPs for population B and 25 SNPs for population C, we randomly selected 5, 10, 15 and 20 SNPs for each population and repeated this 100 times. For each marker set selected, we ran the HapBlock program to identify the total number of blocks, the block boundaries and the Tag SNPs for each block.

Sequential marker selection. Since an SNP could only be a boundary marker in the event that it was in the subset chosen, comparing block boundaries among totally different sets of SNP markers is difficult. In order to further investigate the underlying mechanism explaining why higher density markers usually give rise to more, smaller blocks than is the case for lower density markers, we applied a sequential marker selection method to 46 SNPs on chromosome region 52a from the African-American population. First, we randomly selected

ten SNPs out of the original 46 SNPs to identify block structures. Secondly, we randomly selected another ten SNPs out of the 36 remaining SNPs and combined them with the previously selected 10 SNPs to identify block structures. Then, we randomly selected another 10 SNPs out of the 26 remaining SNPs and combined them with the previously selected 20 SNPs to do the analysis. Lastly, we randomly selected 10 more SNPs out of the 16 remaining SNPs and combined them with the previously selected 30 SNPs to identify block structures. This simulation approach ensured that the lower density marker set is a subset of the higher density marker set. The whole selection process was repeated 100 times. Comparisons of the block boundary results were based on these results.

Block boundary and Tag SNP comparisons

In the comparison of block boundaries, we counted the frequency of each SNP that was used as the starting or ending position of the block boundaries in the results based on 100 randomly selected samples. Comparing Tag SNPs is more complicated than comparing block boundaries because the Tag SNPs are not unique for each block. In other words, there is usually more than one set of Tag SNPs (see Appendix A for a Tag SNP example) in a block. Therefore, to incorporate the multiplicities of the Tag SNPs, for the results from each randomly selected sample, we counted the frequency of each SNP that was selected as a Tag SNP across all Tag SNP sets and divided this frequency by the number of Tag SNP sets in each block and the total number of blocks in the region. Based on the 100 randomly selected samples, we then calculated the mean weighted frequency for each SNP.

Results

Haplotype block partitioning based on the observed data

Using the observed genotype data, region 52a was partitioned into nine blocks with a total of 19 Tag SNPs for the African-Americans (population B) and six blocks with a total of ten Tag SNPs for the Japanese and Chinese (population C). Region 42a, however, was divided into 16 blocks with a total of 33 Tag SNPs for African-Americans and 14 blocks with a total of 22 SNPs for Japanese and Chinese. As with region 31a, both populations had three blocks and six Tag SNPs (see appendix for detailed block information using region 52a as an example).

Inspection of all 51 autosomal regions in the Gabriel *et al.* data set reveals that, in general, chromosomal regions were partitioned into more blocks and had more tag SNPs based on the African-American samples than those based on the Japanese and Chinese samples. In addition, for both populations, the total number of Tag SNPs increases as the number of blocks increases (data not shown).

Impact of sample size

Table 1 summarises the results of the number of blocks when we randomly selected 10, 20, 30 and 40 individuals 100 times from each population. For example, in the upper left

panel of Table 1, column 'ran10' corresponds to the results based on 100 simulated datasets consisting of ten individuals. The sum did not add up to 100 because the HapBlock program we used for block partitioning would tend to fail when

Table 1. Frequency of the number of blocks in which the number of individuals is varied in simulations

Region 52a (Chromosome 22, 237.22 kb)

		African-American				Japanese & Chinese			
		50 individuals, 46 SNPs, 9 blocks				42 individuals, 45 SNPs, 6 blocks			
# blocks		ran10	ran20	ran30	ran40	ran10	ran20	ran30	ran40
10			4	24	36				
9			31	53	49				
8		6	49	23	15				
7		21	14				6	12	
6		55	2				36	75	88
5		17					4	51	19
4							51	13	
3							42		
2									
Sum*		99	100	100	100	98	100	100	100

Region 42a (Chromosome 15, 409.92 kb)

		African-American				Japanese & Chinese			
		50 individuals, 100 SNPs, 16 blocks				42 individuals, 99 SNPs, 14 blocks			
# of blocks		ran10	ran20	ran30	ran40	ran10	ran20	ran30	ran40
19			2	10	5				
18			7	12	19				
17			18	42	53				
16			38	34	23				
15		1	25	2					
14		14	10						
13		24							
12		30					5	13	49
11		22					16	36	27
10		5					33	37	1
9							13	41	14
8							26	5	
7							38		
6							14		
5							1		
Sum*		96	100	100	100	92	100	100	100

Region 31a (Chromosome 9, 181.98 kb)

		African-American				Japanese & Chinese			
		50 individuals, 23 SNPs, 3 blocks				42 individuals, 25 SNPs, 3 blocks			
# of blocks		ran10	ran20	ran30	ran40	ran10	ran20	ran30	ran40
5			1				3	1	
4		2	23	23	29		9	12	4
3		29	63	74	69		61	79	96
2		64	13	3	2		60	23	8
1									
Sum*		95	100	100	100	75	96	100	100

*Sum does not always add up to 100. See results part for detailed explanation.

we had few individuals or few markers included in the sample. Among the 99 simulated samples with HapBlock results, region 52a was partitioned into five blocks 17 times, six blocks 55 times, seven blocks 21 times, and eight blocks six times. If we focus on the trend of modes for each sample size based on 100 simulated samples, it is apparent that the number of blocks generally increases as we include more individuals in the sample. With the original 50 African-Americans, region 52a was partitioned into nine blocks. When we included only ten people, most of the times we obtained six blocks for this region. When we increased the sample size to 20 people, most of the times the region was partitioned into eight blocks. When the sample size grew to 30 and 40, most of the times the region was partitioned into nine blocks, the same as that in the original dataset. Therefore, a minimum of 30 individuals is

needed for this given set of markers to infer the number of blocks.

We also examined the sample size effect on the total number of Tag SNPs associated with block partitioning, and the results are summarised in Table 2. Similar to the results summarised in Table 1, the total number of Tag SNPs increases as the sample size increases. A shorter region with fewer SNPs, such as region 31a, seems to require fewer individuals than a longer region with more SNPs, such as regions 52a and 42a, to identify a similar number of Tag SNPs as the original sample. In fact, the inferred number of blocks and Tag SNPs did not level off in region 42a in either population, indicating that our sample size may not have been adequate to define a set of Tag SNPs for this region. Statistical comparisons based

Table 2. Frequency of the total number of Tag SNPs when the number of individuals is varied in simulations

Region 52a (Chromosome 22, 237.22 kb)

African-American

50 individuals, 46 SNPs,
total 19 Tag SNPs

Japanese & Chinese

42 individuals, 45 SNPs,
total 10 Tag SNPs

Region 42a (Chromosome 15, 409.92 kb)

African-American

50 individuals, 100 SNPs,
total 33 Tag SNPs

Japanese & Chinese

42 individuals, 99 SNPs,
total 22 Tag SNPs

Total # of Tag SNPs	ran10	ran20	ran30	ran40
20		4	5	
19		18	25	
18	13	29	37	
17	37	31	26	
16	1	26	15	7
15	5	16	3	
14	9	6		
13	24	2		
12	33			
11	21			
10	6			
9				
8				
7				
6				
5				
Sum*	99	100	100	100

Total # of Tag SNPs	ran10	ran20	ran30	ran40
38		1	1	2
37		2	2	1
36		2	8	9
35		4	16	22
34		6	21	39
33		18	29	25
32		19	13	2
31		18	8	
30		13	2	
29	2	7		
28	4	8		
27	10	2		
26	9			
25	14			
24	12			
23	19			
22	15			
21	10			
20	1			
19				
18				
17				
16				
15				
14				
13				
12				
11				
10				
9				
8				
7				
6				
5				
4				
3				
Sum*	98	100	100	100

Region 31a (Chromosome 9, 181.98 kb)

African-American

50 individuals, 23 SNPs,
total 6 Tag SNPs

Japanese & Chinese

42 individuals, 25 SNPs,
total 6 Tag SNPs

Total # of Tag SNPs	ran10	ran20	ran30	ran40
8	1	1		
7	2	9	8	11
6	5	32	48	52
5	24	40	40	35
4	51	16	3	2
3	13	2		
Sum*	95	100	100	100

Total # of Tag SNPs	ran10	ran20	ran30	ran40
2		2		
5		5	8	5
2	32	65	95	
6	15	10		
28	26	13		
39	16	4		
75	96	100	100	

*Sum does not always add up to 100. See results part for detailed explanation.

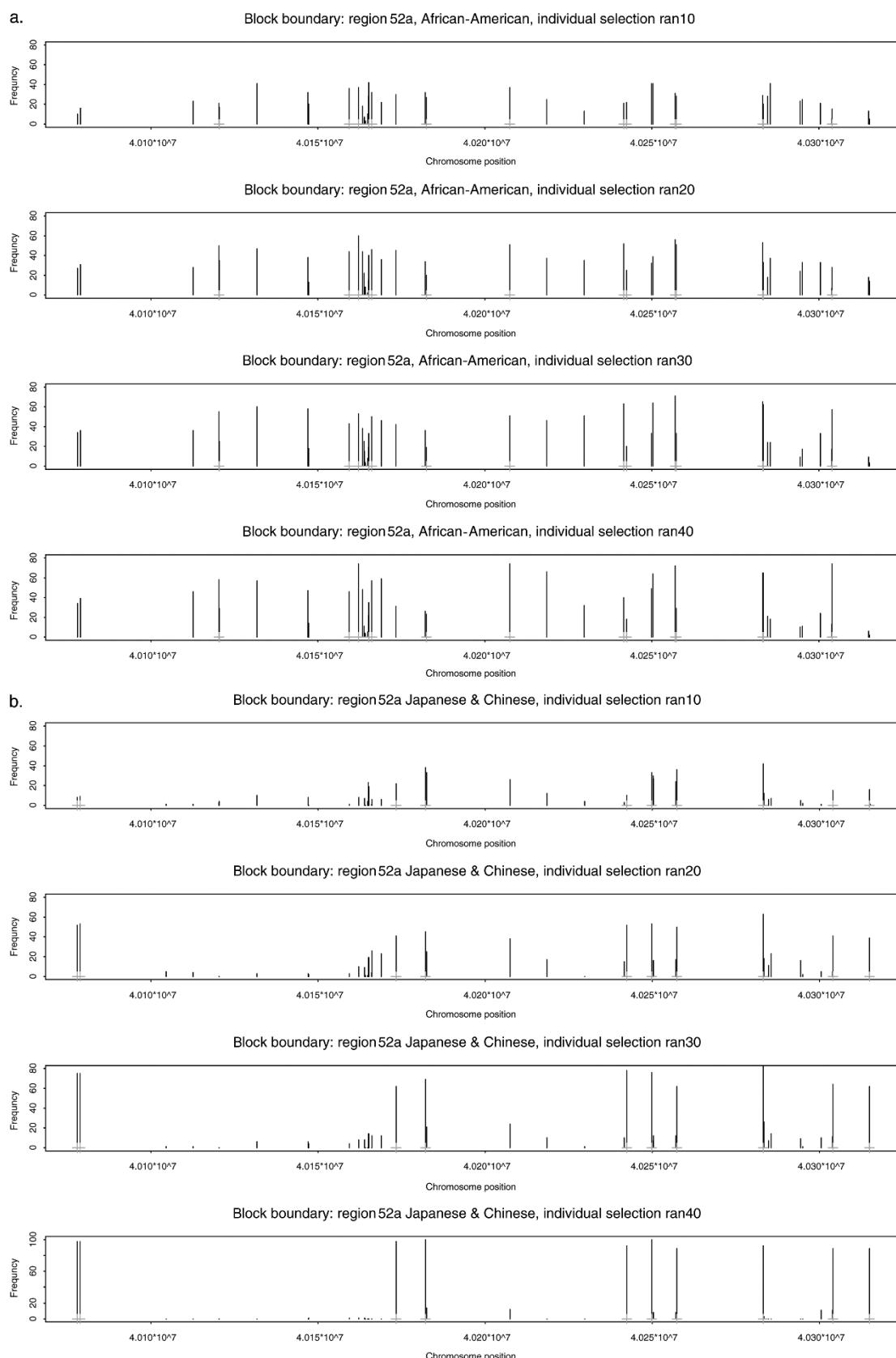


Figure 1. Frequency of each SNP being selected as block boundary against its chromosomal location in individual selection for Region 52a. (a) African-American. (b) Japanese & Chinese. + indicates the position of block boundaries using the original sample.

on t-tests or Wilcoxon tests also indicated that there was a significant difference between the inferred block structures from samples of size 30 and those from samples of size 40 in region 52a.

Using region 52a as an example, Figure 1 summarises the frequency of each SNP being used as block boundary against its chromosomal location across 100 simulated samples with 10, 20, 30 and 40 individuals, respectively. Although block boundaries differed from one sample to another (for samples consisting of the same number of individuals), when we pooled the results of 100 random selections, the overall patterns were very similar for samples of different sizes. The block boundaries in region 52a from the Japanese and Chinese samples were more clear-cut than those from the African-American samples. The high frequency bars matched block boundary positions from those identified in the original 42 Japanese and Chinese people perfectly.

Detailed Tag SNP comparisons are more difficult than block boundary comparisons mainly because Tag SNPs are not unique. Usually there is more than one set of Tag SNPs in a

block (see Appendix A for tag SNP example). In order to examine the impact of sample size on Tag SNP selections, we calculated the weighted frequency of each SNP being selected as a Tag SNP and plotted it against the SNPs in the combined order (see Appendix B for SNPs in the combined order due to differences between SNP sets between the two populations). Figure 2 summarises the results for Tag SNP selections for different sample sizes (10, 20, 30 and 40) and it can clearly be seen that similar sets of Tag SNPs were identified on average across all simulations for different sizes. Comparing these to the Tag SNPs from the original sample of 50 African-Americans, we found that they were almost identical, with the exception of SNP numbers 20 and 45. Both of these had a relatively high frequency of being selected as Tag SNPs using randomly selected samples, but they did not show up in the Tag SNP list using the original sample. In addition, we found that most of the Tag SNPs selected for the Japanese and Chinese population also appeared on the Tag SNP list for the African-American population, but not vice versa, indicating that Tag SNPs for the Japanese and Chinese population

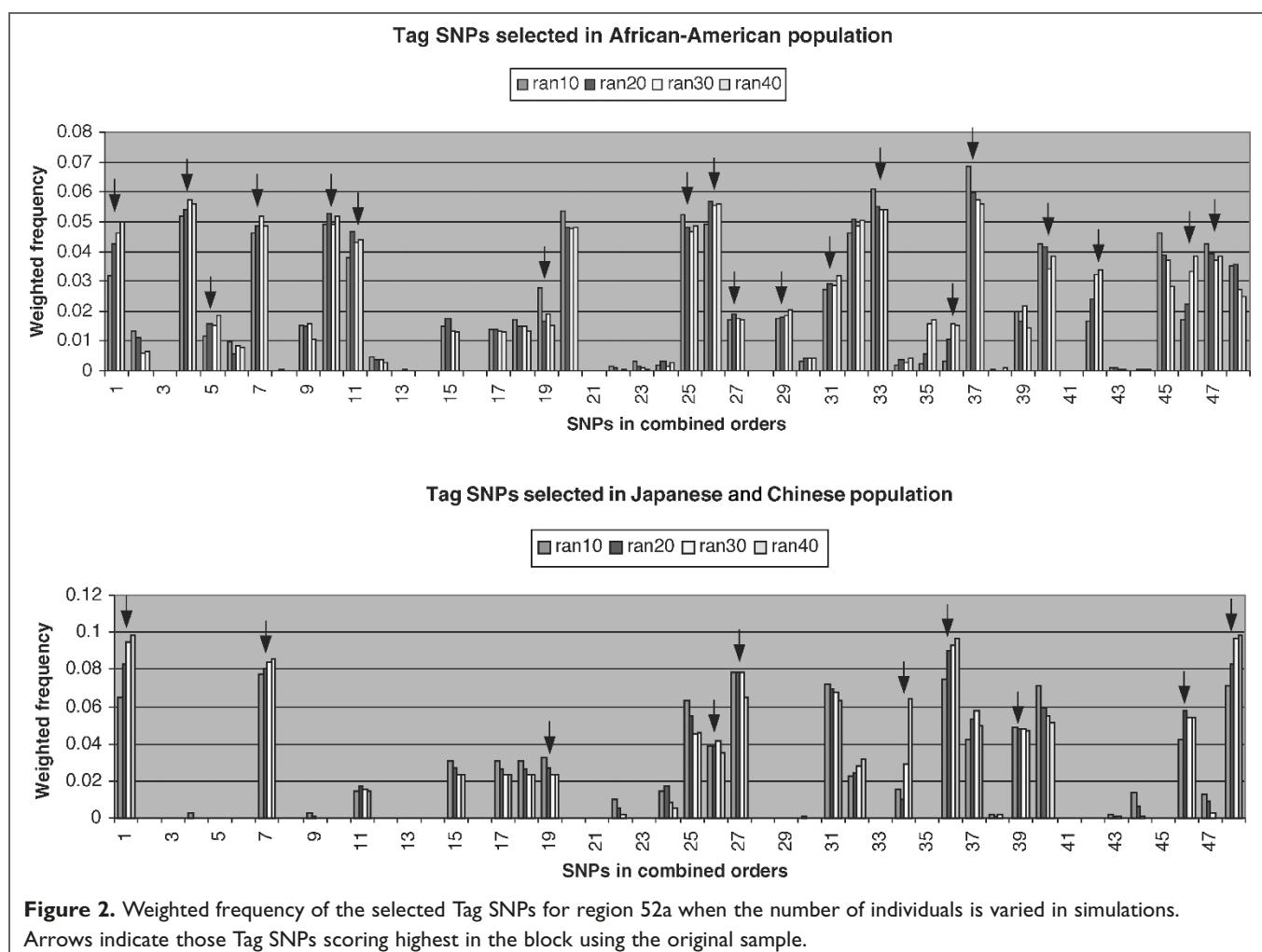


Table 3. Frequency of the number of blocks when the number of markers is varied in simulations

Region 52a (Chromosome 22, 237.22 kb)				
African-American Region		Japanese & Chinese		
50 individuals, 46 SNPs, 9 blocks		42 individuals, 42 SNPs, 6 blocks		
# blocks	ran10	ran20	ran30	ran40
9				11
8				74
7			42	15
6		4	49	
5		40	9	
4	3	54		
3	51	2		
2	45			
Sum*	99	100	100	100

Region 42a (Chromosome 15, 409.92 kb)				
African-American		Japanese & Chinese		
50 individuals, 100 SNPs, 16 blocks		42 individuals, 99 SNPs, 14 blocks		
# blocks	ran20	ran40	ran60	ran80
16				14
15			1	42
14			7	37
13			17	7
12			48	
11		1	25	
10		13	2	
9		40		
8		43		
7	1	3		
6	18			
5	54			
4	27			
3				
2				
Sum*	100	100	100	100

Region 31a (Chromosome 9, 181.98 kb)				
African-American		Japanese & Chinese		
50 individuals, 23 SNPs, 3 blocks		42 individuals, 25 SNPs, 3 blocks		
# blocks	ran10	ran15	ran20	
4			1	
3	2	24	84	
2	72	76	15	
1				
Sum*	74	100	100	

* Sum does not always add up to 100. See results part for detailed explanation.

is largely a subset of those for the African-American population.

Impact of marker selection

Table 3 summarises the results of the number of blocks after we randomly selected: 10, 20, 30 and 40 SNPs for

region 52a; 20, 40, 60 and 80 SNPs for region 42a; and 10, 15 and 20 SNPs for region 31a. Simulated samples consisting of a random selection of five SNPs for region 31a crashed the HapBlock program every time, and therefore no results from this part of the study are shown in Table 3. It is apparent from this Table that as we included more SNP

Table 4. Frequency of the total number of Tag SNPs when the number of markers is varied in simulations**Region 52a** (Chromosome 22, 237.22 kb)**African-American**50 individuals, 46 SNPs,
total 19 Tag SNPs**Japanese & Chinese**42 individuals, 45 SNPs,
total 10 Tag SNPs

Total # of Tag SNPs	ran10	ran20	ran30	ran40
19			1	
18			11	
17			36	
16			41	
15		3	11	
14		14		
13		42		
12		29		
11	5	6		4
10	21	6		5 47
9	29		27	40
8	33		6 45	9
7	2	12	21	19
6	22		4 27	4
5	47		16 29	
4	25		37 16	
3	3		28 1	
2			1	
Sum*	99	100	100	100

Total # of Tag SNPs	ran10	ran20	ran30	ran40
32				3
31				9
30				18
29				28
28				27
27				2 10
26				11 3
25				15 1
24				23 1
23			1	30
22			1	13
21			1	4
20			6	2
19				8
18				
17				
16				
15				
14				
13				
12				
11				
10				
9				
8				
7				
6				
5				
4				
3				
2				
Sum*	86	100	100	100

Region 42a (Chromosome 15, 409.92 kb)**African-American**50 individuals, 100 SNPs,
total 33 Tag SNPs**Japanese & Chinese**42 individuals, 99 SNPs,
total 22 Tag SNPs

Total # of Tag SNPs	ran20	ran40	ran60	ran80
32				3
31				9
30				18
29				28
28				27
27			2	10
26			11	3
25			15	1
24			23	1
23	1	30		
22	1	13		
21	1	4		
20	6	2		
19		8		
18				
17				
16				
15				
14				
13				
12	6			
11	18			
10	31			
9	30			
8				
7				
6				
5				
4				
3				
2				
Sum*	100	100	100	100

Total # of Tag SNPs	ran20	ran40	ran60	ran80
2				
1	15			
1	18			
1	18			
6	25			
13	18			
1	21			
1	27			
5	19			
9	2			
6	29			
12	6			
22				
37				
17				
8				
2				
100	100	100	100	100

*Sum does not always add up to 100. See results part for detailed explanation.

markers in our sample, the number of blocks continued to grow, and there was evidence that the inferred haplotype structures would have continued to change if more markers had been included.

As for the number of Tag SNPs, Table 4 clearly shows that, as we included more SNP markers in our sample, the total number of Tag SNPs also continued to grow, and did not show any sign of stabilisation.

To answer the question of why denser marker sets usually give rise to more, smaller blocks than is the case for sparser marker sets, we studied chromosomal region 52a in the African-American population in detail. Figure 3 shows two representative patterns of how region 52a was partitioned into

blocks using 10, 20, 30 and 40 sequentially-selected SNP markers, as well as the original 46 SNP marker set. Both marker sets of size 10 generated three blocks, with one set consisting of SNPs number 2, 8, 19, 21, 24, 30, 36, 42, 43 and 46, and the other set consisting of SNPs number 5, 6, 11, 15, 22, 23, 24, 25, 40 and 45. The blank space between the blocks is due to the lack of information regarding which block the SNPs belong to. By adding ten more SNPs to both marker sets, the two 20-marker sets generated five blocks, as shown in Figures 3a and 3b. As we included additional SNPs in the marker set within this region, i.e. as we increased the marker density, the number of blocks increased for two reasons. First, the old large blocks at lower densities are often broken into

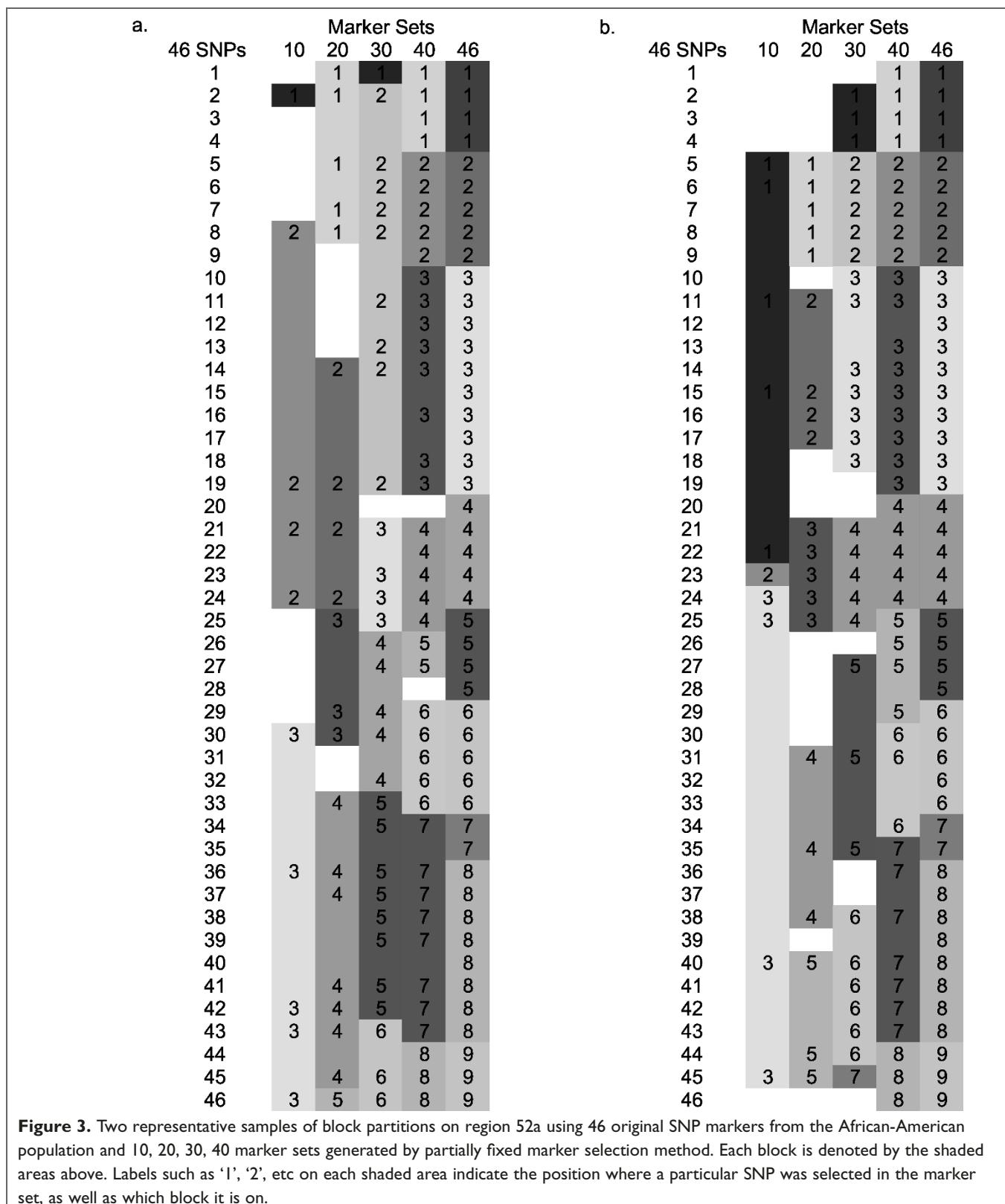


Figure 3. Two representative samples of block partitions on region 52a using 46 original SNP markers from the African-American population and 10, 20, 30, 40 marker sets generated by partially fixed marker selection method. Each block is denoted by the shaded areas above. Labels such as '1', '2', etc on each shaded area indicate the position where a particular SNP was selected in the marker set, as well as which block it is on.

smaller pieces at higher density. For example, in Figure 3a, block 7 in marker set 40 became block 7 and block 8 when two more SNPs (numbers 35 and 40) were added to this

region. Secondly, new blocks emerged from areas where there was a lack of information due to the lack of markers in the smaller marker set, such as block 3 in marker set 20 in

Figure 3a and block 1 in marker set 30 in Figure 3b. The block boundaries were obviously not random but were in fact quite consistent across different marker sets.

Discussion

Our studies have clearly demonstrated that sample size and marker selection have a significant impact on the number of blocks and the total number of Tag SNPs inferred from a population sample. As we include more individuals in our sample, both the number of blocks and the total number of Tag SNPs increase. For a shorter region with fewer SNP markers, like region 31a (181.98 kb, 23 SNPs), 20 people may be adequate to infer the haplotype patterns, while for a longer region with more SNP markers, such as 52a (237.22 kb, 46 SNPs) and 42a (409.92 kb, 100 SNPs), the required sample size may be 30 or more. The minimal sample size needed for a reliable haplotype structure inference clearly depends on the structure of the region being investigated. Although the patterns of block boundary and the set of Tag SNPs selected look very similar on average across all sample sizes, there is more variation from one simulated sample to another when the sample size is small. In addition, the set of Tag SNPs selected in the Japanese and Chinese population seems to be a subset of those in the African-American population.⁸ This observation, however, may be due to the ascertainment of the specific set of markers being examined in the original study.

Our marker selection results demonstrate that the number of blocks and the total number of Tag SNPs increase as more SNP markers are included. In addition, our results indicate that we would need to include more SNP markers in these regions in order to draw a valid conclusion on the number of blocks

and Tag SNPs. The number of SNPs needed for a reliable inference on the haplotype structures may be a function of both the region and the specific population under study.

Another issue to bear in mind is that our haplotypes were inferred from genotype data, not directly observed. Although the accuracy is quite high, greater than 80 per cent,¹⁷ it is likely that the results may differ if different algorithms are used to reconstruct individual haplotypes. In addition, the inaccuracy in haplotype inference may contribute to the observed sample size effect. It should also be noted that the specific set of parameters used in the HapBlock program in our analysis to infer blocks and Tag SNPs does not affect the general patterns for the impact of the sample size and marker selection on the inferred haplotype structures (results not shown).

In summary, our study indicates that sample size and marker selection have a significant impact on the inferred haplotype structures reflected in the haplotype blocks and Tag SNPs. Although haplotype blocks may be an over-simplistic representation of the haplotype structures,¹⁴ we hypothesise that the impact would have been equally significant if we had used other approaches to analysing haplotype structures in the human genome. In order to draw valid conclusions on haplotype block structure, we need a relatively large sample size and a dense marker panel and we need to make adaptive adjustments according to the specific region and specific population to be studied.

Acknowledgments

We thank Dr Kui Zhang for his generous support on the HapBlock program, two reviewers for their constructive comments, and Gabriel and colleagues for making their datasets available. This work was supported in part by NIH grant R01 GM59507 to H.Z.

Appendix A

Region 52a (Chromosome 22, 237.22 kb)

Population B (African-American)[†]

of blocks 9 total # of Tag SNPs 19

BlockID	NumTagSNP	StartPos	EndPos	BlockSize	NumHap
Block_0001	3	1	4	4	100
Block_0002	2	5	9	5	100
Block_0003	2	10	19	10	100
Block_0004	2	20	24	5	100
Block_0005	2	25	28	4	100
Block_0006	2	29	33	5	100
Block_0007	2	34	35	2	100
Block_0008	2	36	43	8	100
Block_0009	2	44	46	3	100

Tag SNP for block_0001

1	4	5	0.95825
1	4	5	0.95825 – *

Tag SNP for block_0005

27	29	0.91095
27	29	0.91095 – *

Tag SNP for block_0002

7	10	0.94339
9	10	0.90594
7	10	0.94339 – *

Tag SNP for block_0006

31	33	0.90816
32	33	0.90614
31	33	0.90816 – *

Tag SNP for block_0003

11	15	0.93446
11	17	0.93107
11	18	0.93234
11	19	0.9346
15	20	0.92013
17	20	0.91561
18	20	0.91455
19	20	0.92754
11	19	0.9346 – *

Tag SNP for block_0007

36	37	
36	37	– *

Tag SNP for block_0008

40	42	0.90181
40	42	0.90181 – *

Tag SNP for block_0009

46	47	0.93096
46	48	0.92839

Tag SNP for block_0004

25	26	0.90927
25	26	0.90927 – *

Tag SNP for block_0009

[†] Tag SNPs are in combined order.
^{*} – | lines indicate the Tag SNPs that scored the highest in each block by the HapBlock program.

Population C (Japanese & Chinese)[†]

of blocks = 6 Total # of TagSNPs = 10

BlockID	NumTagSNP	StartPos	EndPos	BlockSize	NumHap
Block_0001	1	1	1	1	84
Block_0002	2	2	22	21	84
Block_0003	2	23	29	7	84
Block_0004	2	30	34	5	84
Block_0005	2	35	43	9	84
Block_0006	1	44	45	2	84

Tag SNP for block_0001			Tag SNP for block_0004		
1	1.00000		32	36	0.90335
1	1.00000 – *		34	36	0.9073
			34	36	0.9073 – *

Tag SNP for block_0002			Tag SNP for block_0005		
7	15	0.96085	37	39	0.93032
7	17	0.96085	37	40	0.92593
7	18	0.96085	39	46	0.93265
7	19	0.96085	40	46	0.92716
7	15	0.96085 – *	39	46	0.93265 – *

Tag SNP for block_0003			Tag SNP for block_0006		
25	27	0.92191	48	0.95869	
25	31	0.90516	48	0.95869 – *	
26	27	0.92676			
26	31	0.91236			
27	31	0.92645			
26	27	0.92676 – *			

[†]Tag SNPs are in combined order.

*-| lines indicate the Tag SNPs that scored the highest in each block by the HapBlock program.

Appendix B

SNP_ID	COMBINED ORDER	POP_B ORDER	POP_C ORDER	CHROM_POS	POP_B BLOCK	POP_C BLOCK
110924	1	1	1	40077996	Block_0001	Block_0001
110926	2	2	2	40078865	Block_0001	Block_0002
110525	3	NA	3	40104585	NA	Block_0002
110527	4	3	4	40112652	Block_0001	Block_0002
110528	5	4	5	40120338	Block_0001	Block_0002
110529	6	5	6	40120419	Block_0002	Block_0002
3884	7	6	7	40131747	Block_0002	Block_0002
117587	8	7	8	40147031	Block_0002	Block_0002
117590	9	8	9	40147256	Block_0002	Block_0002
91037	10	9	10	40159355	Block_0002	Block_0002
82256	11	10	11	40162170	Block_0003	Block_0002
117575	12	11	NA	40163399	Block_0003	NA
117578	13	12	NA	40163843	Block_0003	NA
3943	14	13	12	40163920	Block_0003	Block_0002
2442	15	14	13	40164108	Block_0003	Block_0002
117580	16	15	14	40164192	Block_0003	Block_0002
117581	17	16	15	40164236	Block_0003	Block_0002
117582	18	17	16	40164840	Block_0003	Block_0002
117583	19	18	17	40165138	Block_0003	Block_0002
37728	20	19	18	40165262	Block_0003	Block_0002
14523	21	NA	19	40166038	NA	Block_0002
82025	22	20	20	40166144	Block_0004	Block_0002
84395	23	21	21	40168971	Block_0004	Block_0002
117586	24	22	22	40173352	Block_0004	Block_0002
117592	25	23	23	40182141	Block_0004	Block_0003
117593	26	24	24	40182498	Block_0004	Block_0003
117596	27	25	25	40207457	Block_0005	Block_0003
26726	28	26	26	40218483	Block_0005	Block_0003
16893	29	27	27	40229786	Block_0005	Block_0003
11692	30	28	28	40241571	Block_0005	Block_0003
117608	31	29	29	40242422	Block_0006	Block_0003
32936	32	30	30	40249849	Block_0006	Block_0004
117566	33	31	31	40250303	Block_0006	Block_0004
44133	34	32	32	40250387	Block_0006	Block_0004
117567	35	33	33	40256951	Block_0006	Block_0004
23139	36	34	34	40257384	Block_0007	Block_0004
118681	37	35	35	40283200	Block_0007	Block_0005
99869	38	36	36	40283420	Block_0008	Block_0005
2584	39	37	37	40284703	Block_0008	Block_0005
118669	40	38	38	40285521	Block_0008	Block_0005
118674	41	39	39	40294440	Block_0008	Block_0005
30109	42	40	40	40295018	Block_0008	Block_0005
118676	43	41	41	40300494	Block_0008	Block_0005
88347	44	42	NA	40303907	Block_0008	Block_0005
118679	45	43	42	40303949	Block_0008	NA
88348	46	44	43	40303993	Block_0009	Block_0005
3742	47	45	44	40314969	Block_0009	Block_0006
54	48	46	45	40315218	Block_0009	Block_0006

References

1. Kruglyak, L. and Nickerson, D.A. (2001), 'Variation is the spice of life', *Nat. Genet.* Vol. 27, pp. 234–236.
2. Sachidanandam, R., Eeissman, D., Schmidt, S.C. *et al.* (2001), 'A map of human genome sequence variation containing 1.42 million single nucleotide polymorphisms', *Nature* Vol. 409, pp. 928–933.
3. Venter, J.C., Adams, N.D., Myers, E.W. *et al.* (2001), 'The sequence of the human genome', *Science* Vol. 291, pp. 1304–1351.
4. Stephens, J.C., Schneider, J.A., Tanguay, D.A. *et al.* (2001), 'Haplotype variation and linkage disequilibrium in 313 human genes', *Science* Vol. 293, pp. 489–493.
5. Daly, M.J., Rioux, J.D., Schaffnel, S.F. *et al.* (2001), 'High-resolution haplotype structure in the human genome', *Nat. Genet.* Vol. 29, pp. 229–232.
6. Patil, N., Berno, A.J., Hurd, D.A. *et al.* (2001), 'Blocks of limited haplotype diversity revealed by high-resolution scanning of human chromosome', *Science* Vol. 294, pp. 1719–1723.
7. Dawson, E., Abecasis, G.R., Bumpstead, S. *et al.* (2002), 'A first-generation linkage disequilibrium map of human chromosome 22', *Nature* Vol. 418, pp. 544–548.
8. Gabriel, S.B., Schaffnel, S.F., Nguyen, H. *et al.* (2002), 'The structure of haplotype blocks in the human genome', *Science* Vol. 296, pp. 2225–2229.
9. Jeffreys, A.J., Kauppi, L. and Neumann, R. (2001), 'Intensely punctate meiotic recombination in the class II region of the major histocompatibility complex', *Nat. Genet.* Vol. 29, pp. 217–222.
10. Zhang, K., Calabrese, P., Nordborg, M. and Sun, F. (2002), 'Haplotype block structure and its applications to association studies: Power and study designs', *Am. J. Hum. Genet.* Vol. 71, pp. 1386–1394.
11. Reich, D.E., Cargill, M., Bolk, S. *et al.* (2001), 'Linkage disequilibrium in the human genome', *Nature* Vol. 411, pp. 199–204.
12. Zhang, K., Deng, H., Chen, T. *et al.* (2002), 'A dynamic programming algorithm for haplotype block partitioning', *Proc. Natl. Acad. Sci. USA* Vol. 99, pp. 7335–7339.
13. Wang, N., Akey, J.M., Zhang, K. *et al.* (2002), 'Distribution of recombination crossovers and the origin of haplotype blocks: The interplay of population history, recombination, and mutation', *Am. J. Hum. Genet.* Vol. 71, pp. 1227–1234.
14. Wall, J.D. and Pritchard, J.K. (2003), 'Haplotype blocks and linkage disequilibrium in the human genome', *Nat. Rev. Genet.* Vol. 4, pp. 587–597.
15. Wall, J.D. and Pritchard, J.K. (2003), 'Assessing the performance of the haplotype block model of linkage disequilibrium', *Am. J. Hum. Genet.* Vol. 73, pp. 502–515.
16. Johnson, G.C., Esposito, L., Barratt, B.J. *et al.* (2001), 'Haplotype tagging for the identification of common disease genes', *Nat. Genet.* Vol. 29, pp. 233–237.
17. Stephens, M., Smith, N.J. and Donnelly, P. (2001), 'A new statistical method for haplotype reconstruction from population data', *Am. J. Hum. Genet.* Vol. 68, pp. 978–989.