

# Update on the olfactory receptor (OR) gene superfamily

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## Abstract

The olfactory receptor gene (*OR*) superfamily is the largest in the human genome. The superfamily contains 390 putatively functional genes and 465 pseudogenes arranged into 18 gene families and 300 subfamilies. Even members within the same subfamily are often located on different chromosomes. *OR* genes are located on all autosomes except chromosome 20, plus the X chromosome but not the Y chromosome. The gene:pseudogene ratio is lowest in human, higher in chimpanzee and highest in rat and mouse — most likely reflecting the greater need of olfaction for survival in the rodent than in the human. The *OR* genes undergo allelic exclusion, each sensory neurone expressing usually only one odourant receptor allele; the mechanism by which this phenomenon is regulated is not yet understood. The nomenclature system (based on evolutionary divergence of genes into families and subfamilies of the *OR* gene superfamily) has been designed similarly to that originally used for the *CYP* gene superfamily.

**Keywords:** classification of gene families and subfamilies, *OR* gene superfamily, *CYP* gene superfamily, nasal olfactory neurone, olfaction, olfactory receptor gene superfamily, allelic exclusion, opossum genome, platypus genome

## Introduction

Before 1980, the names of genes and classification of their encoded proteins were highly variable and non-systematic — especially to anyone slightly outside a particular field or to a new graduate student entering the field. Professor Margaret Oakley Dayhoff was a pioneer in attempting to create order out of chaos in the naming of genes and gene families by means of computerised protein alignments.<sup>1</sup> She was widely recognised as the founder in this new field of gene/protein classification, before her untimely death in 1983.

Cytochrome P450 (*CYP*) genes are conveniently arranged into families and subfamilies based on the percentage amino acid sequence identity.<sup>2–7</sup> Enzymes that share approximately  $\geq 40$  per cent

identity are assigned to a particular family designated by an Arabic numeral, whereas those sharing approximately  $\geq 55$  per cent identity are grouped into a particular subfamily designated by a letter. For example, the sterol 27-hydroxylase enzyme and the 25-hydroxy-vitamin D<sub>3</sub> 1 $\alpha$ -hydroxylase enzyme are both assigned to the *CYP27* family because they share  $>40$  per cent sequence identity. Furthermore, the sterol 27-hydroxylase is assigned to the *CYP27* 'A' subfamily and the 25-hydroxy-vitamin D<sub>3</sub> 1 $\alpha$ -hydroxylase to the *CYP27* 'B' subfamily because their protein sequences are  $<55$  per cent identical. If an additional enzyme were to be discovered that shared  $>55$  per cent identity with the sterol 27-hydroxylase, then it would be named *CYP27A2*. If an additional enzyme were to be discovered that

shared <55 per cent but >40 per cent identity with the sterol 27-hydroxylase as well as the 25-hydroxy-vitamin D<sub>3</sub> 1 $\alpha$ -hydroxylase, then it would be named CYP27C1. The development and application of this delightfully logical system of nomenclature to the genes of many animals, plants and bacteria<sup>8</sup> has eliminated the confusion that often had plagued the naming of gene families and superfamilies. Subsequently, this 'divergent evolution' nomenclature system was adopted for several hundred other gene families and superfamilies — including the olfactory receptor superfamily.

## Background and history

Vertebrate olfactory receptor (*OR*) genes represent a category of G-protein-coupled receptors (GPCRs) that contain seven transmembrane  $\alpha$ -helical domains and function in the reception of innumerable odour molecules in the environment.<sup>9</sup> The *OR* gene superfamily is the largest in vertebrate genomes.<sup>10–13</sup> The genomic architecture of mammalian *OR* gene clusters shows an ancient evolutionary origin, preceding the marsupial–eutherian split; species-specific evolution has further shaped the different *OR* gene families, by means of both gains and losses of complete clusters, as well as expansion and contraction of existing clusters.<sup>11</sup>

This dynamic flexibility is also reflected among individual humans; examining 51 candidate *OR* genes on DNA chips in 189 ethnically diverse subjects, a striking amount of population diversity was found.<sup>14</sup> Segregating pseudogenes (SPGs) are genes that segregate in populations between intact genes and pseudogenes — due to a disruptive single nucleotide polymorphism (SNP). A range of 16–24 functional *OR* genes was found, just in this study alone, indicating that the *OR* gene superfamily is among the most pronounced examples of functional population diversity in the human genome.<sup>14</sup> Copy number variations (CNVs), another type of polymorphism, are also highly prevalent among human *OR* genes.<sup>15,16</sup> All these genomic events are evidence of a relatively recent process, whereby the extreme diminution of a

functional repertoire in humans has occurred — a process which is presumably still ongoing.

For most mammalian species, the ability to detect millions of different odourants is critical to their survival. Based on recent *OR* gene mining data in the platypus, opossum, cow and dog genomes — compared with that in the rat, mouse, macaque and human genomes<sup>13</sup> — we are now certain that there has been a substantial expansion of the *OR* gene superfamily since the mammalian radiation ~100 million years ago.

The evolutionary change in the number of *OR* genes in insects is not nearly as extensive as that in mammals. *Drosophila melanogaster* has a relatively

**Table 1.** Summary of the olfactory receptor (*OR*) gene superfamily (18 families)

Family	No. of subfamilies	No. of functional genes	No. of pseudogenes
1	21	28	11
2	41	68	45
3	3	4	2
4	21	51	78
5	49	47	65
6	21	30	21
7	9	11	100
8	18	24	24
9	12	9	14
10	29	35	30
11	11	8	15
12	1	2	1
13	11	12	10
14	6	6	1
51	21	23	21
52	22	26	23
55	1	0	1
56	2	6	3
Totals	299	390	465

**Table 2.** Summary of OR genes in families 1 to 4

Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo
OR1A	2	0	OR2I	0	1	OR3A	4	0
OR1B	1	0	OR2J	2	2	OR3B	0	1
OR1C	1	0	OR2K	1	0	OR3D	0	1
OR1D	3	1	OR2L	5	3			
OR1E	2	1	OR2M	5	1	OR4A	4	27
OR1F	2	1	OR2N	0	1	OR4B	1	1
OR1G	1	0	OR2P	0	1	OR4C	10	11
OR1H	0	1	OR2Q	0	1	OR4D	7	3
OR1I	1	0	OR2R	0	1	OR4E	1	1
OR1J	3	0	OR2S	1	1	OR4F	9	7
OR1K	1	0	OR2T	16	1	OR4G	0	6
OR1L	5	0	OR2U	0	2	OR4H	0	2
OR1M	1	1	OR2V	2	0	OR4K	8	7
OR1N	2	0	OR2W	3	3	OR4L	1	0
OR1P	0	1	OR2X	0	1	OR4M	2	0
OR1Q	1	0	OR2Y	1	0	OR4N	3	2
OR1R	0	1	OR2Z	1	0	OR4P	1	1
OR1S	2	0	OR2AD	0	1	OR4Q	1	2
OR1X	0	2	OR2AE	1	0	OR4R	0	3
OR1AA	0	1	OR2AF	0	1	OR4S	2	0
OR1AB	0	1	OR2AG	2	0	OR4T	0	1
			OR2AH	0	1	OR4U	0	1
			OR2AI	0	1	OR4V	0	1
OR2A	9	6	OR2AJ	1	0	OR4W	0	1
OR2B	4	3	OR2AK	1	0	OR4X	2	2
OR2C	2	0	OR2AL	0	1			
OR2D	2	0	OR2AM	0	1			
OR2E	0	1	OR2AO	0	1			
OR2F	2	0	OR2AP	1	0			
OR2G	3	1	OR2AS	0	2			
OR2H	2	2	OR2AT	1	2			

Family OR1 genes are located on chromosomes 9, 17, 19, 11, 16, 5, 1, 6 and X.

Family OR2 genes are located on chromosomes 1, 6, 7, 11, 5, 9, 12, 16, 19 and X.

Family OR3 genes are located on chromosomes 17, 1 and X.

Family OR4 genes are located on chromosomes 11, 14, 15, 1, 19, 17, 18, 21, 6, 4, 5, 8 and X.

small receptor repertoire of 62 odourant receptors.<sup>17</sup> A comparison of 12 *Drosophila* species, encompassing ~60 million years of divergence, shows that the number of functional OR genes has remained fairly stable.<sup>18</sup> *Caenorhabditis elegans* has a highly developed chemosensory system, which enables it to detect a wide variety of volatile (olfactory) and water-soluble (gustatory) cues associated with food, danger or other animals; between 500 and 1,000 different GPCRs are expressed in chemosensory neurones, and these may be supplemented by alternative sensory pathways as well.<sup>19</sup> The vertebrate OR gene repertoire has thus evolved from a subset of ancestral genes in the fly and worm.

There appear to be three important periods in the evolution of the vertebrate olfactory system, as evidenced by comparative genomics: (1) the adaptation to land in amphibian ancestors; (2) the decline of olfaction in primates; and (3) the delineation of putative pheromone receptors concurrent with rodent speciation.<sup>20</sup> The gene:pseudogene ratio is lowest in human, higher in chimpanzee and highest in rat and mouse. This most likely reflects the necessity of olfaction for survival — more so in the rodent than in the human.

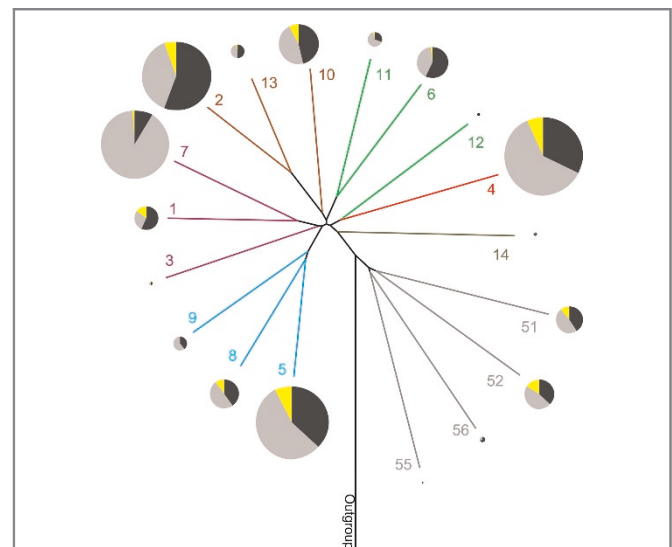
Whereas the chicken, platypus and primate genomes carry <400 functional OR genes, the opossum and rodent genomes, not surprisingly, contain between 1,000 and 1,210 functional OR genes.<sup>11,13</sup> Curiously, however, it is difficult to explain why the cow genome, with 970 functional OR genes, shows more than the dog genome, with ~811 functional OR genes, when dogs are considered to have such a keen sense of olfaction.<sup>13</sup> Thus, the number of OR genes in a species does not appear to be directly related to the environmental 'requirement' or to lifestyle.

## Current bioinformatics about the OR gene superfamily

The OR gene superfamily comprises 18 gene families and 300 subfamilies (Table 1). Presently, there are 390 putatively functional (protein-coding)

OR genes and 465 OR pseudogenes located in multiple clusters of varying sizes scattered throughout all autosomes except chromosome (Chr) 20, and on the X but not the Y chromosome.<sup>21–23</sup>

The members of each subfamily have been placed therein because of divergent evolution, as described above. These subfamilies differ from CYP subfamilies, in that individual members within one subfamily are often located on two or more different chromosomes. The OR2T (Table 2) subfamily contains 16 functional genes — more than in any other subfamily. Evolutionary divergence of each of the 18 gene families is illustrated in Figure 1.



**Figure 1.** A phylogenetic analysis of one representative from each family of the human OR gene repertoire. In this tree, one can see the following: (1) a general guideline for how the different families relate to one another (although this is very general, and the branching is not always this well defined); (2) the numbers near each branch denote the OR family number; (3) each pie chart size is scaled to represent the number of the OR genes inside that family (black = functional genes, grey = pseudogenes, yellow = segregating pseudogenes [SPGs]). SPGs are genes that segregate in populations between intact genes and pseudogenes — due to a disruptive SNP.<sup>34</sup> This disruptive mutation can introduce a stop codon, or alter a highly conserved amino acid that is important for proper function of the protein. In Tables 1–5, the SPGs are counted as 'functional genes' or 'pseudogenes', according to the Human Genome Project version. Additional information can be found at the HORDE database (<http://bioportal.weizmann.ac.il/HORDE/>).

**Table 3.** Summary of OR genes in families 5 to 8

Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo
OR5A	2	0	OR5BC	0	1	OR7A	3	8
OR5B	5	4	OR5BD	0	1	OR7C	2	0
OR5C	1	0	OR5BE	0	1	OR7D	2	1
OR5D	4	4	OR5BH	0	1	OR7E	1	85
OR5E	0	1	OR5BJ	0	1	OR7G	3	1
OR5F	1	1	OR5BK	0	1	OR7H	0	2
OR5G	0	4	OR5BL	0	1	OR7K	0	1
OR5H	5	5	OR5BM	0	1	OR7L	0	1
OR5I	1	0	OR5BN	0	2	OR7M	0	1
OR5J	1	2	OR5BP	0	1			
OR5K	4	0	OR5BQ	0	1	OR8A	1	2
OR5L	2	0	OR5BR	0	1	OR8B	5	6
OR5M	6	8	OR5BS	0	1	OR8C	0	1
OR5P	2	2	OR5BT	0	1	OR8D	3	0
OR5R	1	0				OR8F	0	1
OR5S	0	1	OR6A	1	0	OR8G	2	3
OR5T	3	0	OR6B	3	0	OR8H	3	0
OR5V	1	0	OR6C	11	8	OR8I	1	2
OR5W	1	1	OR6D	0	1	OR8J	2	1
OR5AC	1	2	OR6E	0	1	OR8K	3	2
OR5AH	0	1	OR6F	1	0	OR8L	0	1
OR5AK	1	3	OR6J	1	0	OR8Q	0	1
OR5AL	0	2	OR6K	3	3	OR8R	0	1
OR5AM	0	1	OR6L	0	2	OR8S	1	0
OR5AN	1	1	OR6M	1	2	OR8T	0	1
OR5AO	0	1	OR6N	2	0	OR8U	3	0
OR5AP	1	1	OR6P	1	0	OR8V	0	1
OR5AQ	0	1	OR6Q	1	0	OR8X	0	1
OR5AR	1	0	OR6R	0	2			
OR5AS	1	0	OR6S	1	0			
OR5AU	1	0	OR6T	1	0			

Continued

**Table 3.** Continued

Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo
OR5AW	0	1	OR6U	0	1			
OR5AZ	0	1	OR6V	1	0			
OR5BA	0	1	OR6W	0	1			
OR5BB	0	1	OR6X	1	0			
			OR6Y	1	0			

Family OR5 genes are located on chromosomes 11, 3, 12, 6, X, 14, 19, 2, 4 and 9.

Family OR6 genes are located on chromosomes 12, 1, 11, 14, 10, 7, 2 and 8.

Family OR7 genes are located on chromosomes 19, 11, 3, 8, 4, 13, 2, 12, 7, 10, 14, 9, 21, 5 and X.

Family OR8 genes are located only on chromosomes 11 and 12.

**Table 4.** Summary of OR genes in families 9 to 13

Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo
OR9A	2	2	OR10A	6	0	OR11A	1	0
OR9G	3	2	OR10B	0	1	OR11G	1	1
OR9H	0	1	OR10C	1	0	OR11H	5	4
OR9I	1	2	OR10D	0	4	OR11I	0	1
OR9K	1	1	OR10G	7	2	OR11J	0	3
OR9L	0	1	OR10H	5	0	OR11K	0	2
OR9M	0	1	OR10J	3	6	OR11L	1	0
OR9N	0	1	OR10K	2	0	OR11M	0	1
OR9P	0	1	OR10N	0	1	OR11N	0	1
OR9Q	2	0	OR10P	1	0	OR11P	0	1
OR9R	0	1	OR10Q	1	1	OR11Q	0	1
OR9S	0	1	OR10R	1	2			
			OR10S	1	0	OR12D	1	2
			OR10T	1	1			
			OR10U	0	1	OR13A	1	0
			OR10V	1	2	OR13C	6	3
			OR10W	1	0	OR13D	1	2
			OR10X	1	0	OR13E	0	1
			OR10Y	0	1	OR13F	1	0
			OR10Z	1	0	OR13G	1	0
			OR10AA	0	1	OR13H	1	0
			OR10AB	0	1	OR13I	0	1

Continued

**Table 4.** Continued

Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo
			OR10AC	0	1	OR13J	1	0
			OR10AD	1	0	OR13K	0	1
			OR10AE	0	2	OR13Z	0	2
			OR10AF	0	1			
			OR10AG	1	0	OR14A	2	0
			OR10AH	0	1	OR14C	1	0
			OR10AK	0	1	OR14I	1	0
						OR14J	1	0
						OR14K	1	0
						OR14L	0	1

Family OR9 genes are located on chromosomes 11, 7, 12, 1 and 2.

Family OR10 genes are located on chromosomes 11, 1, 19, 12, 14, 7 and 6.

Family OR11 genes are located on chromosomes 14, 15, 1, X, 6, 12 and 22.

Family OR12 genes are located only on chromosome 6.

Family OR13 genes are located on chromosomes 9, 1, X and 10.

Family OR14 genes are located only on chromosome 1.

**Table 5.** Summary of OR genes in families 51, 52, 55 and 56

Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo
OR51A	3	7	OR52A	3	0	OR55B	0	1
OR51B	4	2	OR52B	3	3			
OR51C	0	2	OR52D	1	0	OR56A	4	1
OR51D	1	0	OR52E	5	3	OR56B	2	2
OR51E	2	0	OR52H	1	1			
OR51F	2	3	OR52I	2	0			
OR51G	2	0	OR52J	1	2			
OR51H	0	2	OR52K	2	1			
OR51I	2	0	OR52L	1	1			
OR51J	1	0	OR52M	1	1			
OR51K	0	1	OR52N	4	1			
OR51L	1	0	OR52P	0	2			
OR51M	1	0	OR52Q	0	1			
OR51N	0	1	OR52R	1	0			
OR51P	0	1	OR52S	0	1			
OR51Q	1	0	OR52T	0	1			

Continued

Table 5. Continued

Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo	Subfamily	Genes	Pseudo
OR51R	0	1	OR52U	0	1			
OR51S	1	0	OR52V	0	1			
OR51T	1	0	OR52W	1	0			
OR51V	1	0	OR52X	0	1			
OR51AB	0	1	OR52Y	0	1			
			OR52X	0	1			

The OR51, OR52, OR55 and OR56 genes are located only on chromosome 11.

Note that, in many instances, some subfamilies contain only a single gene or only a single pseudogene (Tables 2–5). In fact, the *OR7E* subfamily has only one functional gene, and all the other 85 members are pseudogenes (Table 3). The *OR7E* subfamily is the largest subfamily in the human *OR* gene repertoire, and probably has expanded in the human genome through a series of segmental gene duplication events.<sup>24</sup> The newly described human *OR14* gene family (Table 4) was realised after analysis of the platypus and opossum *OR* gene repertoires. This analysis revealed that six human *OR* functional genes and one *OR* pseudogene (which previously had been classified within the *OR5* family) are actually derived from a distinct platypus *OR* gene family.<sup>11,25</sup> The evolutionary divergence of the *OR14* gene family is shown in Figure 2.

The ‘shotgun’ splattering of *OR* genes throughout the human genome must have happened before speciation of *Homo sapiens* and the development of its 22 autosomes plus the X and Y chromosomes; this can be inferred from the high conservation of the *OR* genes’ genomic organisation among marsupial and eutherian mammals,<sup>11</sup> and the phylogenetic analysis of the platypus *OR* gene repertoire — by comparison with that in mammals.<sup>13,25</sup> In contrast to this *OR* gene arrangement would be the establishment of the *CYP* gene subfamilies, which arose as syntenic clusters of members within a single chromosomal segment. This finding suggests that gene duplication events within *CYP* subfamilies occurred after mammalian speciation and development of the autosomes and sex chromosomes.

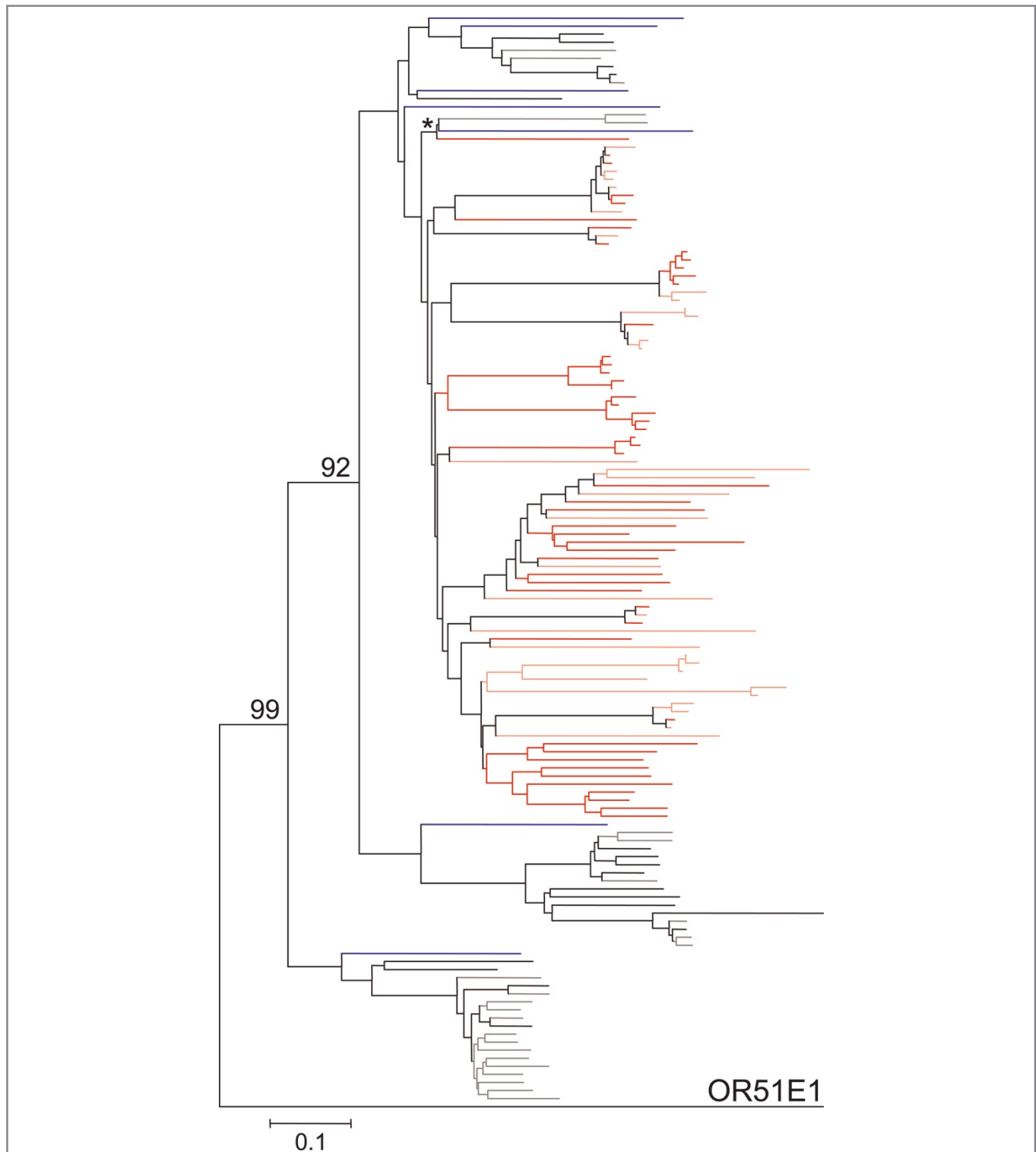
The two largest *OR* gene clusters are located on Chr 11, with 38 functional genes (51 per cent of total) on 11q (Cluster 11@5.0) and 44 functional genes (45 per cent) on 11p (Cluster 11@55.6). These genes are predominantly in *OR* families 51, 52, 55 and 56 (Table 5). Immersed within these two clusters are dozens of other non-*OR*-related genes. This intrusion of other non-*OR*-related genes can also be seen in all other *OR* gene clusters throughout the genome.

### Future directions: Additional subsets of sensory reception genes and identification of ligands

A recently appreciated discovery in olfaction is the unique specialisation of sensory neurones, such that each individual sensory neurone is stochastically chosen to express usually only one odourant receptor allele. This mechanism of ‘allelic exclusion’, by which mutually exclusive expression of odourant receptor genes is regulated, remains unclear at present.<sup>20,26,27</sup>

The vomeronasal-1 receptor genes (*VN1R*) also encode GPCRs and, while they encode odourant receptors, they are evolutionarily distinct<sup>20</sup> from the very large *OR* gene superfamily. There are five *VN1R* genes and nine *VN1R* pseudogenes. The *VN1R1*, *VN1R2* and *VN1R4* genes and *VN1R6P* pseudogene are located at Chr 19q13.42; the *VN1R10P*, *VN1R11P*, *VN1R12P*, *VN1R13P* and *VN1R14P* pseudogenes are located on Chr 6p21; *VN1R7P* and *VN1R8P* are on Chr 21p11.2;





**Figure 2.** A phylogenetic analysis of platypus, opossum and human *OR* genes for the new family 14 only. Opossum = black for intact genes, grey for pseudogenes. Platypus = red for intact genes, pink for pseudogenes. Human = blue. *OR14* is an expansion of three ancestral *OR* gene subfamilies (A, B and C); the expansion, in both platypus and opossum, took place after speciation, whereas only one branch shows an orthologous relationship between platypus and human (marked with \*). The tree was generated with MEGA4, using the nearest-neighbour-joining algorithm, and distances with the Poisson correction model. Bootstrap units are also indicated.<sup>34</sup> This tree is grounded with the *OR51E1* gene. Only genes with no more than two frame disruptions were considered in the analysis.

*VN1R3* is alone on Chr 16p11.2; *VN1R5* is alone on Chr 1q44; *VN1R9P* is alone on Chr 22.<sup>28</sup>

At the present time, information about the ligands for mammalian *OR* genes is very limited. The smell of lemons (limonene), the perception of a floral or woody smell (acetophenone)<sup>29</sup> and the ability to smell isovaleric acid<sup>30</sup> have been mapped in the mouse to two specific genomic loci on Chr 4 (*Iva1*) and Chr 6 (*Iva2*). In humans, isovaleric acid was found to be highly associated with the *OR11H7P* segregating pseudogene, which is not syntenic with either *Iva1* or *Iva2*.<sup>31</sup> Another recent study found that human *OR7D4* is selectively activated *in vitro* by androstenone; interestingly, this study found that two non-synonymous SNPs account for a significant proportion of the variance in smell perception of androstenone.<sup>32</sup>

Members of the gustatory receptor (*Gr*) gene family in *Drosophila* are expressed in chemosensory neurones and are known to mediate the perception of sugars, bitter substrates, carbon dioxide and pheromones. Intriguingly, some of these *Gr* genes have now been shown to be expressed in abdominal multi-dendritic neurones, hygrosensitive neurones of the arista, peripheral proprioceptive neurones in the legs, neurones in the larval and adult brain, and oenocytes.<sup>33</sup> Along these same lines, we and others have observed several *OR* genes being significantly up- or downregulated in the liver or kidney of knockout mouse lines — that is, in tissues not normally known to be involved in olfaction. It is therefore tempting to speculate that the receptors encoded by *OR* genes, as well as by *Gr* genes, might participate in the roles of detecting endogenous ligands.

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## References

- Dayhoff, M.O., Barker, W.C. and Hunt, L.T. (1983), 'Establishing homologies in protein sequences', *Methods Enzymol.* Vol. 91, pp. 524–545.
- Nebert, D.W., Adesnik, M., Coon, M.J. *et al.* (1987), 'The P450 gene superfamily. Recommended nomenclature', *DNA Vol.* 6, pp. 1–11.
- Nebert, D.W. and Gonzalez, F.J. (1987), 'P450 genes: Structure, evolution, and regulation', *Annu. Rev. Biochem.* Vol. 56, pp. 945–993.
- Nebert, D.W., Nelson, D.R., Adesnik, M. *et al.* (1989), 'The P450 superfamily: Updated listing of all genes and recommended nomenclature for the chromosomal loci', *DNA Vol.* 8, pp. 1–13.
- Nebert, D.W., Nelson, D.R., Coon, M.J. *et al.* (1991), 'The P450 superfamily: Update on new sequences, gene mapping, and recommended nomenclature', *DNA Cell Biol.* Vol. 10, pp. 1–14.
- Nelson, D.R., Kamataki, T., Waxman, D.J. *et al.* (1993), 'The P450 superfamily: Update on new sequences, gene mapping, accession numbers, early trivial names of enzymes, and nomenclature', *DNA Cell Biol.* Vol. 12, pp. 1–51.
- Nelson, D.R., Koymans, L., Kamataki, T. *et al.* (1996), 'P450 superfamily: Update on new sequences, gene mapping, accession numbers and nomenclature', *Pharmacogenetics* Vol. 6, pp. 1–42.
- <http://drnelson.utmem.edu/CytochromeP450.html>.
- Buck, L. and Axel, R. (1991), 'A novel multigene family may encode odorant receptors: A molecular basis for odor recognition', *Cell* Vol. 65, pp. 175–187.
- Gaillard, I., Rouquier, S. and Giorgi, D. (2004), 'Olfactory receptors', *Cell Mol. Life Sci.* Vol. 61, pp. 456–469.
- Aloni, R., Olender, T. and Lancet, D. (2006), 'Ancient genomic architecture for mammalian olfactory receptor clusters', *Genome Biol.* Vol. 7, p. R88.
- Henion, T.R. and Schwarting, G.A. (2007), 'Patterning the developing and regenerating olfactory system', *J. Cell Physiol.* Vol. 210, pp. 290–297.
- Niimura, Y. and Nei, M. (2007), 'Extensive gains and losses of olfactory receptor genes in mammalian evolution', *PLoS ONE* Vol. 2, p. e708.
- Menashe, I., Man, O., Lancet, D. and Gilad, Y. (2003), 'Different noses for different people', *Nat. Genet.* Vol. 34, pp. 143–144.
- Nguyen, D.Q., Webber, C. and Ponting, C.P. (2006), 'Bias of selection on human copy-number variants', *PLoS Genet.* Vol. 2, p. e20.
- Hasin, Y., Olender, T., Khen, M. *et al.* (2008), 'High resolution copy-number variation map reflects human olfactory receptor diversity and evolution', *PLoS Genet.* *in press*.
- Smith, D.P. (2007), 'Odor and pheromone detection in *Drosophila melanogaster*', *Pflüger's Arch.* Vol. 454, pp. 749–758.
- Nozawa, M. and Nei, M. (2007), 'Evolutionary dynamics of olfactory receptor genes in *Drosophila* species', *Proc. Natl. Acad. Sci. USA* Vol. 104, pp. 7122–7127.
- Bargmann, C.I. (2006), 'Chemosensation in *C. elegans*', *Wormbook*, October 25, pp. 1–29.
- Kambere, M.B. and Lane, R.P. (2007), 'Co-regulation of a large and rapidly evolving repertoire of odorant receptor genes', *BMC Neurosci.* Vol. 8 (Suppl. 3), p. S2.
- <http://biportal.weizmann.ac.il/HORDE/>.
- Safra, M., Chalifa-Caspi, V., Shmueli, O. *et al.* (2003), 'Human gene-centric databases at the Weizmann Institute of Science: GeneCards, UDB, CroW 21 and HORDE', *Nucleic Acids Res.* Vol. 31, pp. 142–146.
- Olender, T., Feldmesser, E., Atarot, T. *et al.* (2004), 'The olfactory receptor universe — From whole genome analysis to structure and evolution', *Genet. Mol. Res.* Vol. 3, pp. 545–553.
- Yue, Y. and Haaf, T. (2006), '7E olfactory receptor gene clusters and evolutionary chromosome rearrangements', *Cytogenet. Genome Res.* Vol. 112, pp. 6–10.
- Warren, W.C., Hillier, L.W., Marshall-Graves, J.A. *et al.* (2008), 'Genome analysis of the platypus reveals unique signatures of evolution', *Nature* Vol. 453, pp. 175–183.
- Serizawa, S., Miyamichi, K., Nakatani, H. *et al.* (2003), 'Negative feedback regulation ensures the one receptor-one olfactory neuron rule in mouse', *Science* Vol. 302, pp. 2088–2094.
- Rodriguez, I. (2007), 'Odorant and pheromone receptor gene regulation in vertebrates', *Curr. Opin. Genet. Dev.* Vol. 17, pp. 465–470.
- <http://www.genenames.org/>.
- Malnic, B., Godfrey, P.A. and Buck, L.B. (2004), 'The human olfactory receptor gene family', *Proc. Natl. Acad. Sci. USA* Vol. 101, pp. 2584–2589.

30. Griff, I.C. and Reed, R.R. (1995), 'The genetic basis for specific anosmia to isovaleric acid in the mouse', *Cell* Vol. 83, pp. 407–414.
31. Menashe, I., Abaffy, T., Hasin, Y. *et al.* (2007), 'Genetic elucidation of human hyperosmia to isovaleric acid', *PLoS Biol.* Vol. 5, p. e284.
32. Keller, A., Zhuang, H., Chi, Q. *et al.* (2007), 'Genetic variation in a human odorant receptor alters odour perception', *Nature* Vol. 449, pp. 468–472.
33. Thorne, N. and Amrein, H. (2008), 'Atypical expression of *Drosophila* gustatory receptor genes in sensory and central neurons', *J. Comp. Neurol.* Vol. 506, pp. 548–568.
34. Tamura, K., Dudley, J., Nei, M. and Kumar, S. (2007), 'MEGA4: Molecular evolutionary genetics analysis (MEGA) software version 4.0', *Mol. Biol. Evol.* Vol. 24, pp. 1596–1599.