

LETTER TO THE EDITOR

Open Access



Letter to the editor for “Update of the human and mouse Fanconi anemia genes”

Daniel W. Nebert^{1,2*}, Hongbin Dong³, Elspeth A. Bruford⁴, David C. Thompson⁵, Hans Joenje⁶ and Vasilis Vasiliou^{3*}

After our review of the “Update of the human and mouse Fanconi anemia genes” had been published [1], a report appeared [2] that we believe contains additional important and clarifying information relevant to our discussion of the “Fanconi anemia pathway” and cross-talk with other DNA-repair pathways.

In Fig. 1, which is a modified version of the Figure 2 cartoon in our recent review [1], we had shown that, in response to upstream DNA damage signaling (such as phosphorylation by ATR/ATM) in the “FA/BRCA pathway,” the FA core complex comprises at least 11 proteins. These include FANCA (A), FANCB (B), FANCC (C), FANCE (E), FANCF (F), FANCG (G), FANCM (M), and FANCL (L) proteins, plus three FAAP proteins (FAAP20, FAAP24, and FAAP100). This core complex binds UBE2T (T) by way of FANCL; the resultant complex then activates FANCD2/I dimers by means of mono-ubiquitination, and we had written that this is an essential prerequisite for repairing DNA interstrand cross-links.

The activated FANCD2/I (D2/I) complex then translocates to DNA damage sites and recruits downstream FA effector proteins. These include BRCA1 (S), BRCA2 (D1), RAD51 (R), BRIP1 (J), PALB2 (N), RAD51C (O), SLX4 (P), and ERCC4 (Q), plus other DNA-repair molecules (including FA-associated nuclease-1 (*FANL1*)), as illustrated as the orange ellipse at the far right in our Fig. 1 diagram [1]. This large complex then migrates to the site of the lesion to repair the DNA damage.

It had been presumed that FAN1, also required for interstrand cross-link repair, is recruited by ubiquitinated FANCD2/I (light green ellipse in Fig. 1) [1].

However, Lachaud and coworkers, using FAN1 nuclease-deficient mice, recently demonstrated [2] that recruitment of FAN1 by ubiquitinated FANCD2/I is not essential for interstrand cross-link repair. As an alternative, FAN1 recruitment and activity restrain DNA replication fork progression. This restraint, in turn, holds in check any chromosomal abnormalities from occurring, when the DNA replication forks stall. And this pause can happen—even in the absence of interstrand cross-links.

Consequently, recruitment of FAN1 by ubiquitinated FANCD2 may be regarded as facilitating the processing of stalled forks during DNA replication. Although this checkpoint process is essential for genome stability and improved overall well-being of the cell, it might better be described as not absolutely necessary. Therefore, a slightly revised *figure* is proposed here, in which FAN1 can be either bound or not bound from the remainder of this complex (comprising D2/I, S, D1, R, J, N, O, P, Q, and other DNA repair molecules). This *revised diagram* now takes into account the latest data published by Lachaud et al. [2].

Future experiments using other knockout mice will clarify this pathway further. Given the recent advances with CRISPR/*Cas9* methodology, knockout mouse lines—or cells in culture—can be efficiently created for all components shown in the *figure* herein, and then all permutations can be examined, one-by-one.

Acknowledgements

This work was supported in part by the NIH Grants AA022057 and AA021724 (WV), U41HG003345 (EAB), P30 ES006096 (DWN), and the Wellcome Trust grant 099129/Z/12/Z (EAB).

Authors' contributions

DWN drafted the manuscript. EAB, DCT, HJ, and WV edited the manuscript. All authors read and approved the final manuscript.

Competing interests

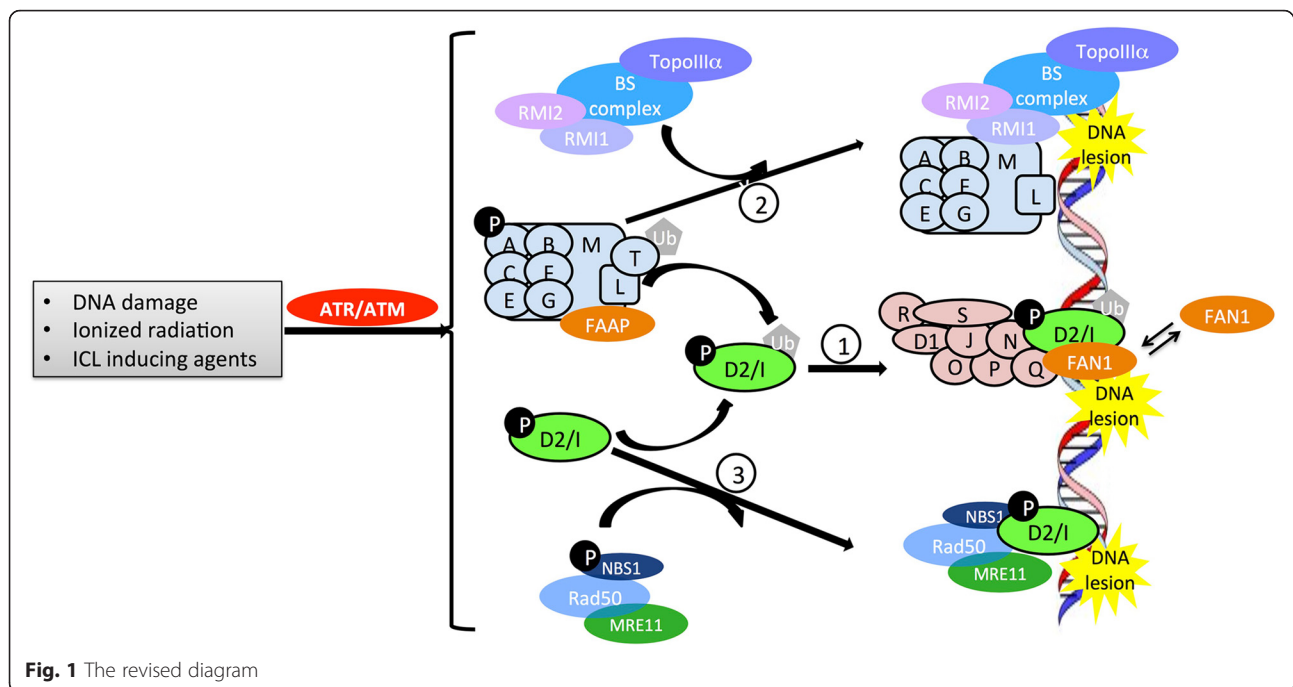
The authors declare that they have no competing interests.

* Correspondence: dan.nebert@uc.edu; vasilis.vasiliou@yale.edu

¹Department of Pediatrics and Molecular Developmental Biology, Division of Human Genetics, Children's Hospital Medical Center, Cincinnati, OH 45229-2899, USA

³Department of Environmental Health Sciences, Yale School of Public Health, 60 College St., New Haven, CT 06250, USA

Full list of author information is available at the end of the article



Author details

¹Department of Pediatrics and Molecular Developmental Biology, Division of Human Genetics, Children's Hospital Medical Center, Cincinnati, OH 45229-2899, USA. ²Department of Environmental Health and Center for Environmental Genetics, University Cincinnati Medical Center, P.O. Box 0056, Cincinnati, OH 45267-0056, USA. ³Department of Environmental Health Sciences, Yale School of Public Health, 60 College St., New Haven, CT 06250, USA. ⁴HUGO Gene Nomenclature Committee (HGNC), European Bioinformatics Institute-European Molecular Biology Laboratory (EMBL-EBI), HinxtonCB10 1SDUK. ⁵Department of Clinical Practice, University of Colorado Denver, Aurora, CO 80045, USA. ⁶Department of Clinical Genetics and the Cancer Center, Amsterdam/VUmc Institute for Cancer and Immunology, VU University Medical Center, NL-1081 BT, Amsterdam, The Netherlands.

Received: 14 May 2016 Accepted: 14 June 2016

Published online: 04 July 2016

References

- Dong H, Nebert DW, Bruford EA, Thompson DC, Joenje H, Vasiliou V. Update of the human and mouse Fanconi anemia genes. *Hum Genomics*. 2015;9(1):32.
- Lachaud C, Moreno A, Marchesi F, Toth R, Blow JJ, Rouse J. Ubiquitinated FANCD2 recruits FAN1 to stalled replication forks to prevent genome instability. *Science*. 2016;351(6275):846–9.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

